

# Tools for Mapping the Circuits of Intelligence

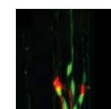
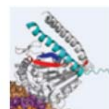
---

**Ed Boyden**

Synthetic Neurobiology Group  
MIT



Biological Engineering



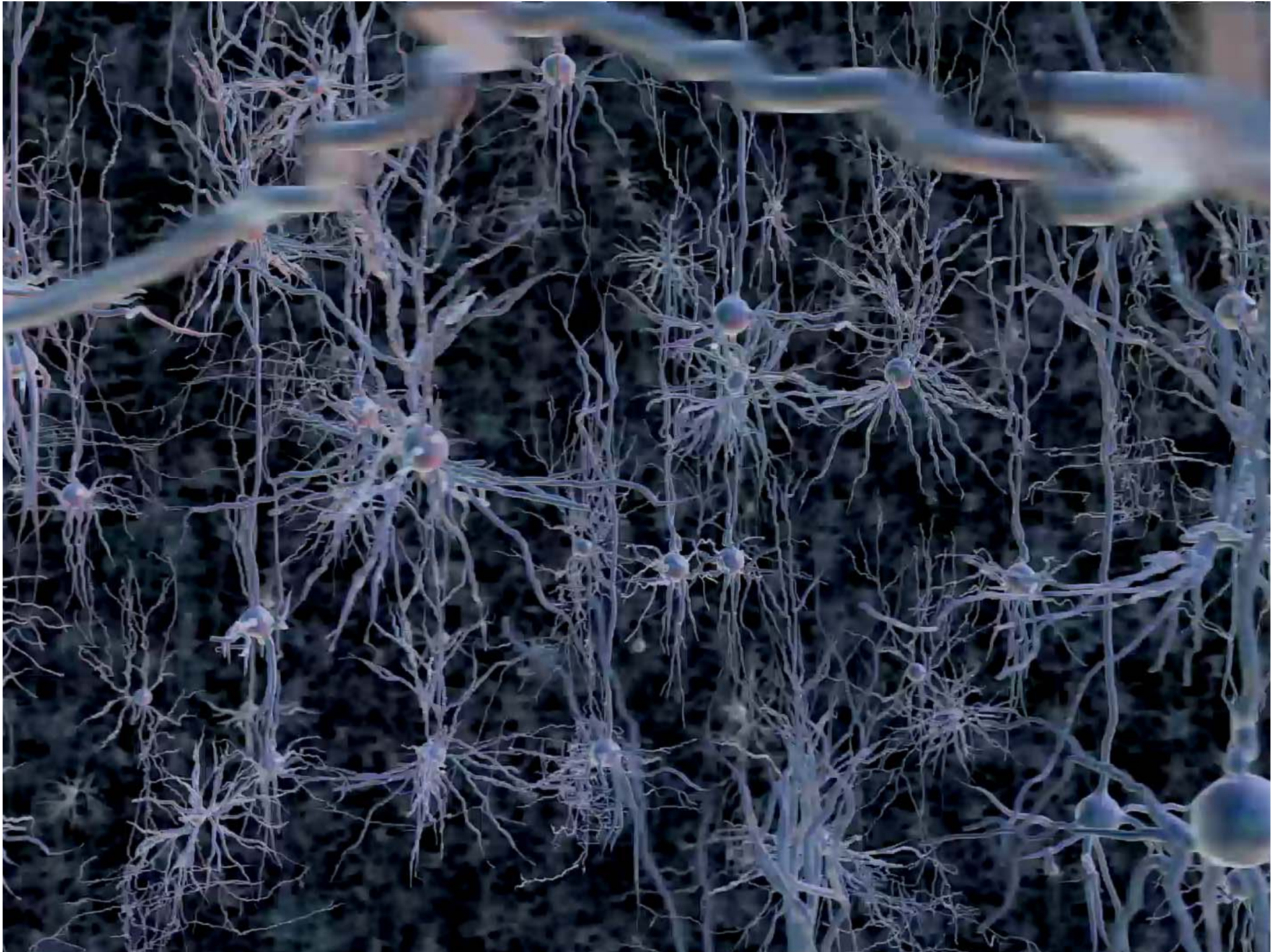
MIT CENTER FOR NEUROBIOLOGICAL ENGINEERING

**Understanding the brain is a problem of fundamental difficulty, but also of scale**

Organized at nanoscale, but spanning centimeters ( $10^7$  range)

Computing with millisecond timescale events, but spanning years ( $10^{12}$  range)







# Principles of neuroengineering design:

Work **backwards** from properties of the brain,

Survey the entire scope of **engineering** possibility,

**Invent** technologies for analyzing and engineering the brain

**Map**

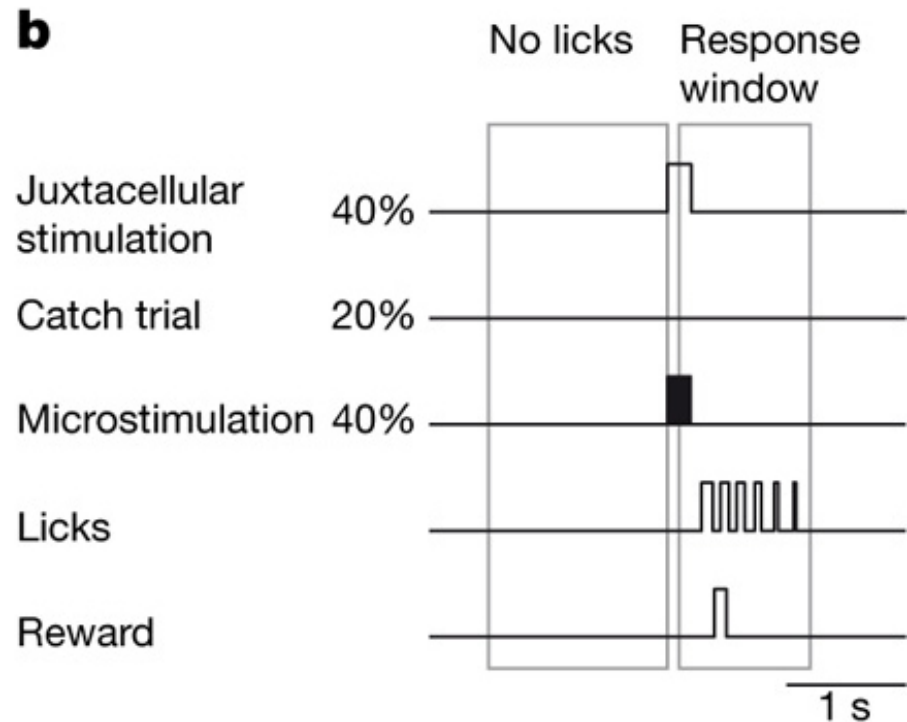
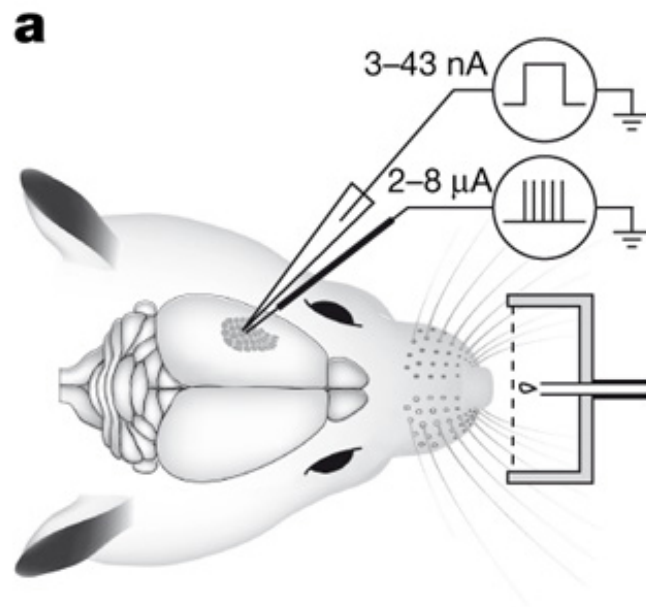
**Record**

**Control**

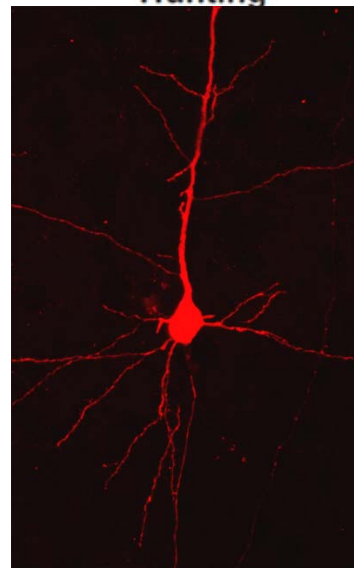
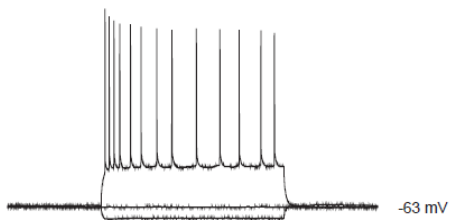
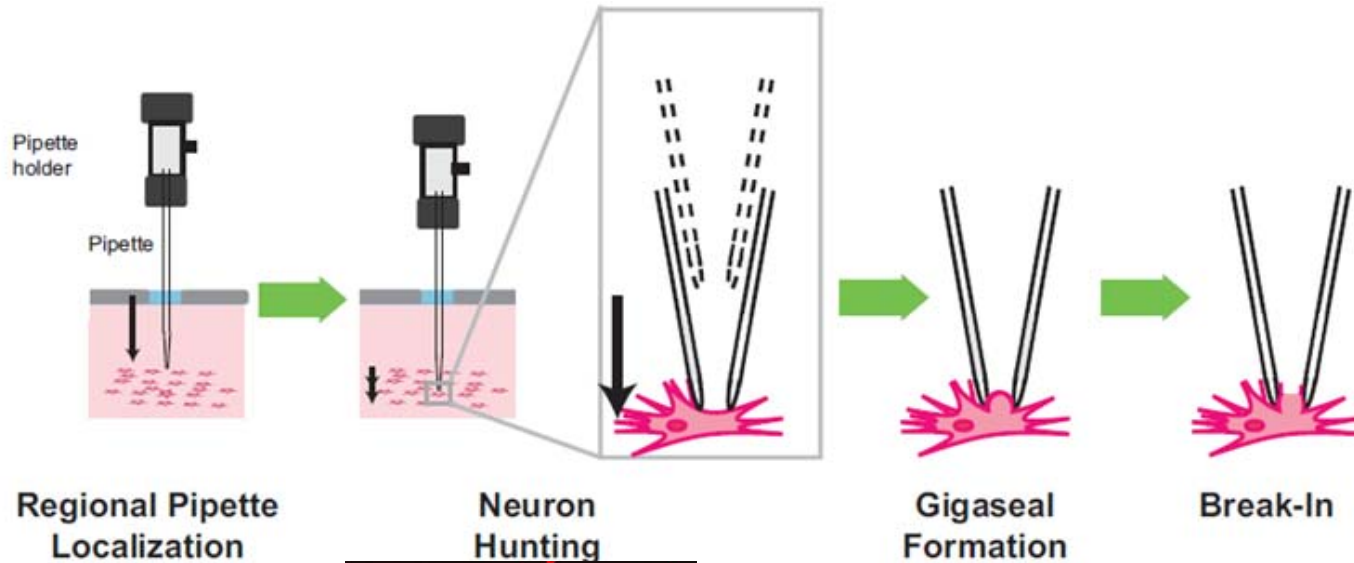
**Build**



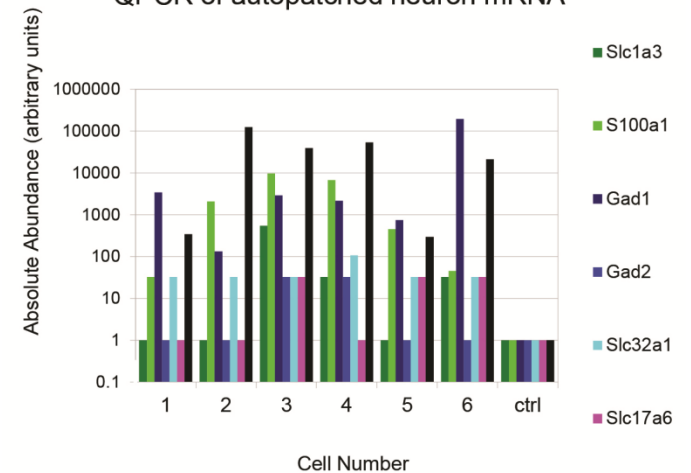
# How does a single neuron compute?



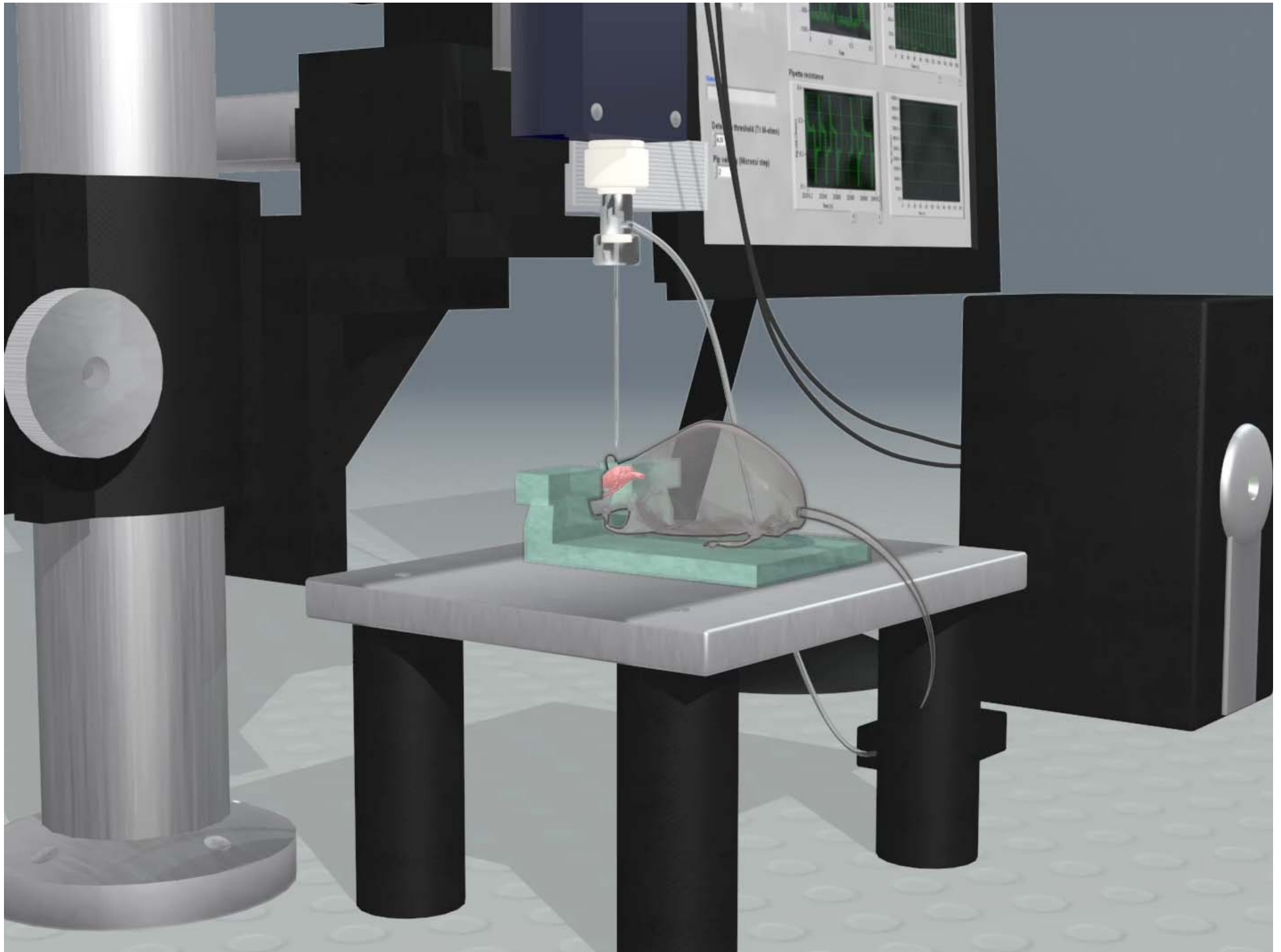
# Whole cell patch clamp: enables simultaneous measurement of electrophysiology, morphology, and gene expression in single cells in living brain



QPCR of autopatched neuron mRNA

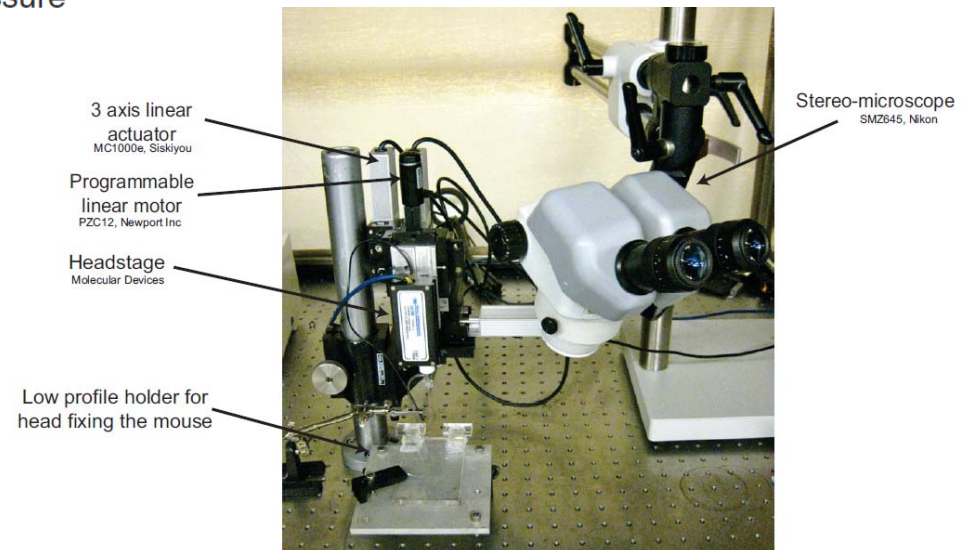
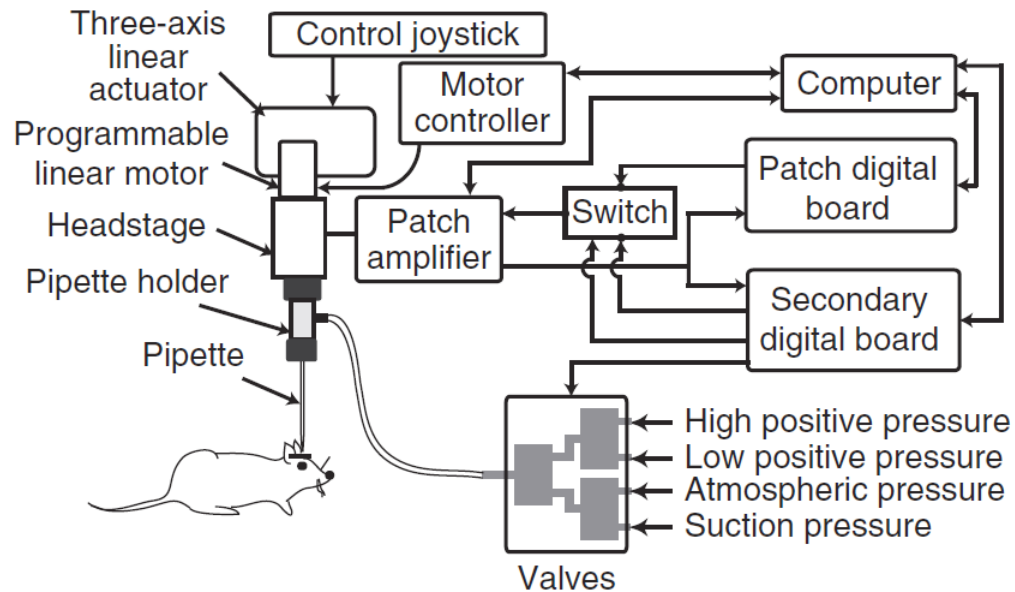


Kodandaramaiah et al. (2012) *Nature Methods* 9:585–587.



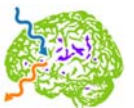


# A robot that can automatically patch clamp neurons in living brain

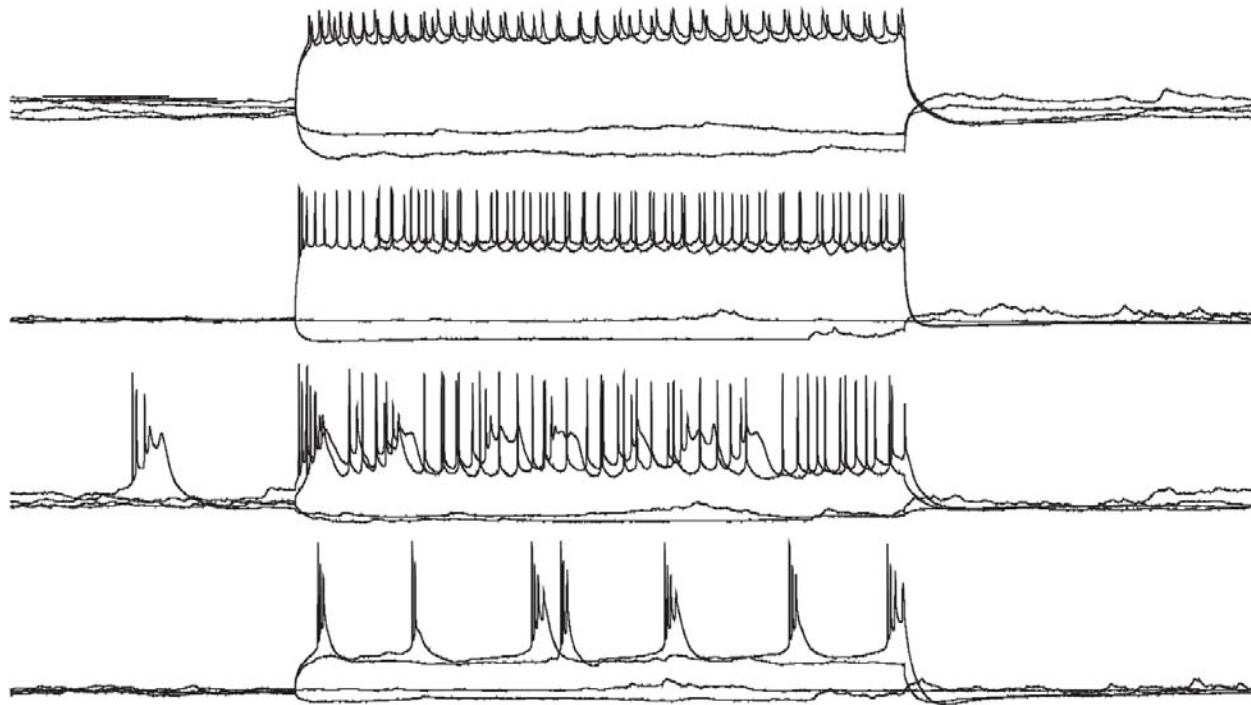


Kodandaramaiah et al. (2012) *Nature Methods* 9:585–587.

Commercialized by Neuromatic Devices, Inc. (ESB has no financial affiliation)



# Robotic quad patching in living mouse brain



Suhasa Kodandaramaiah, Francisco Flores, Emery Brown, Craig Forest



# How do neurons work together in entire circuits?



# C. elegans

(Sulston and Horvitz, 1977)

“There are **302** neurons in the nervous system of *C. elegans*; this number is invariant between animals. Each neuron has a **unique combination of properties, such as morphology, connectivity and position, so that every neuron may be given a unique label.**

Groups of neurons that differ from each other only in position have been assigned to classes. There are **118 classes that have been made using these criteria, the class sizes ranging from 1 to 13.**

Thus *C. elegans* has a rich variety of neuron types in spite of having only a small total complement of neurons. This is in marked contrast to structures such as the mammalian cerebellum, which contains more than  $10^{10}$  neurons (Braitenberg & Atwood 1958) and yet has only five classes of component neuron (Eccles *et al.* 1967).”

White..Brenner, Phil. Trans. Royal Soc. London. Series B, Biol. Sci. Vol.314, Issue 1165 (Nov 12, 1986), 1-340

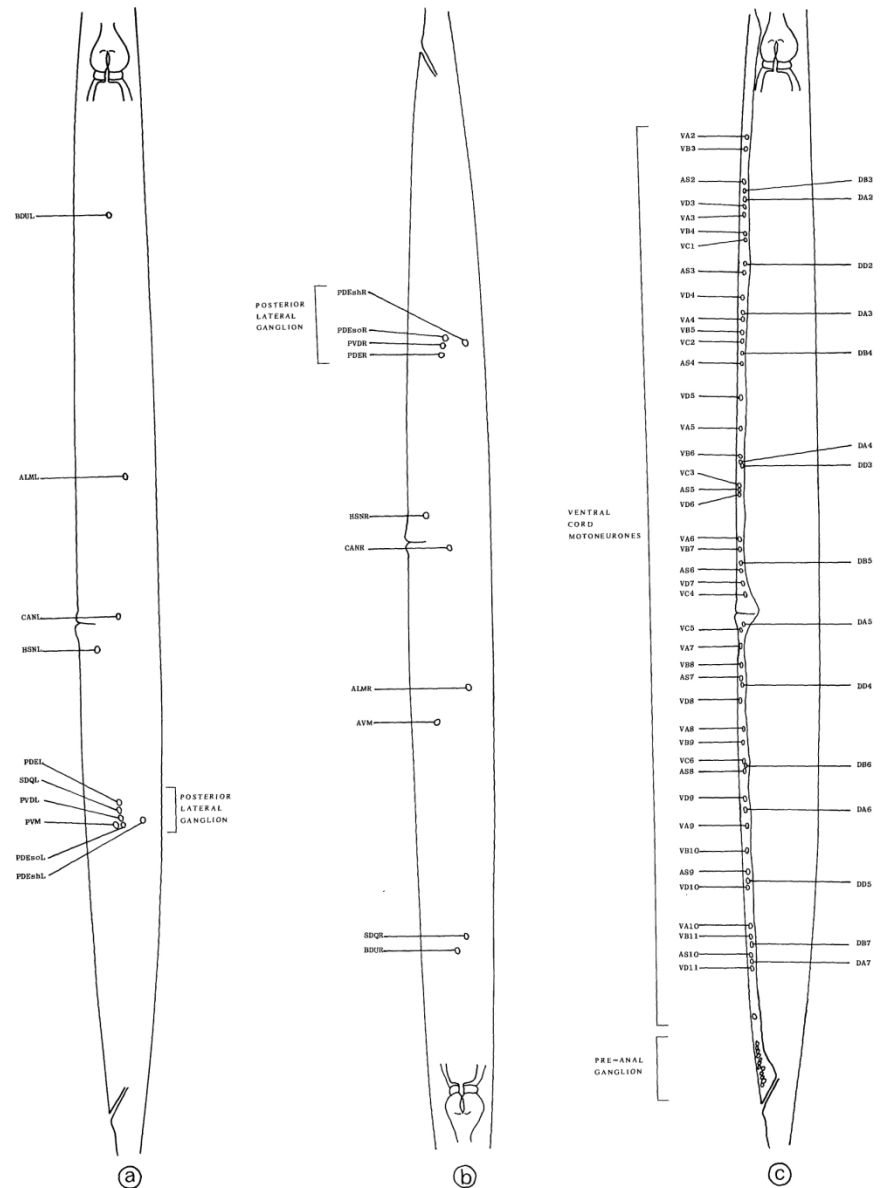


FIGURE 4. For description see opposite.

# C. elegans

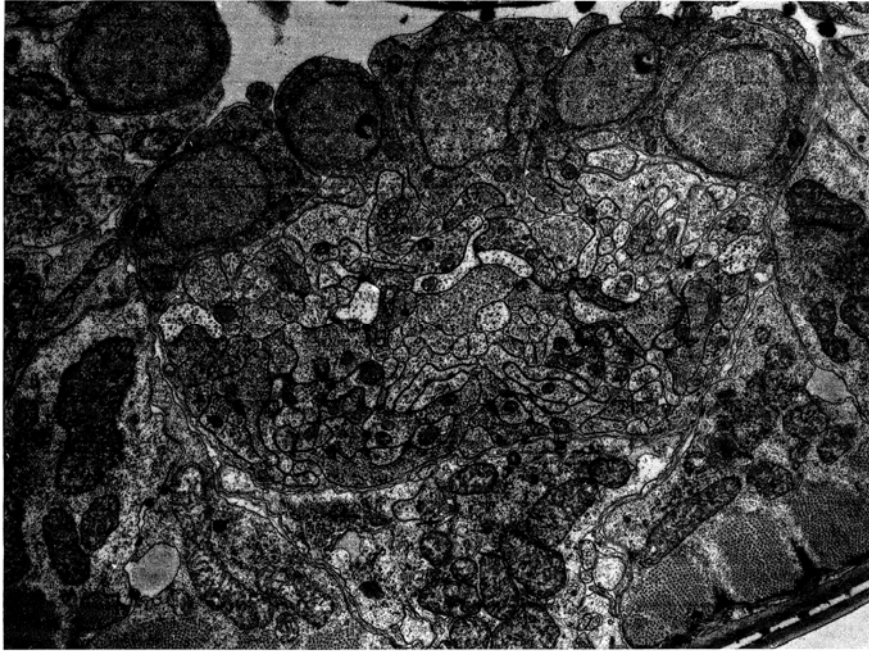
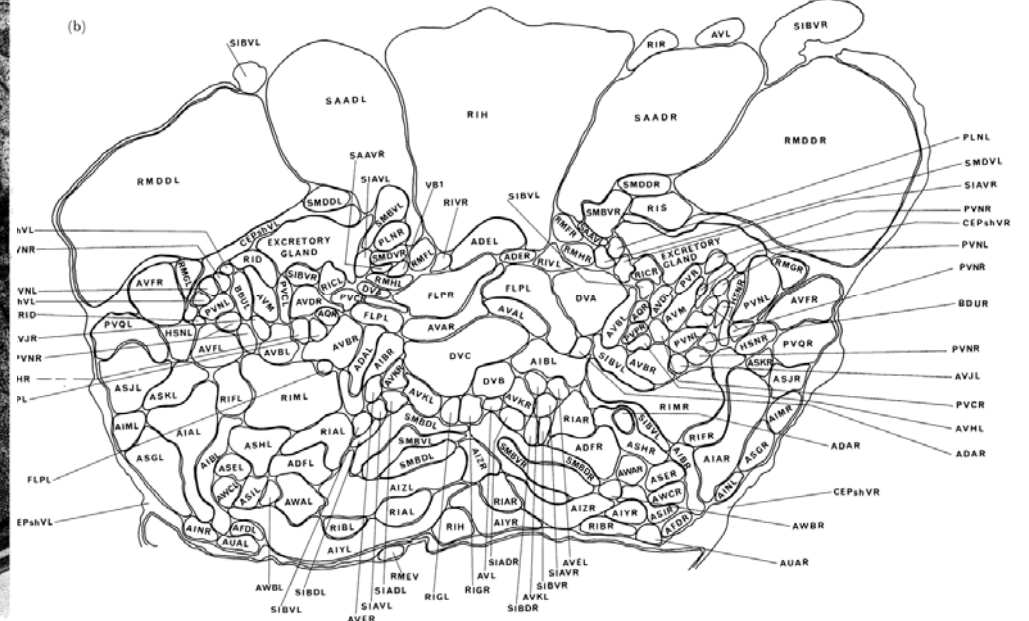


Figure 16(a). For description see opposite.



[http://www.wormatlas.org/MoW\\_built0.92/nervous\\_system.html](http://www.wormatlas.org/MoW_built0.92/nervous_system.html)

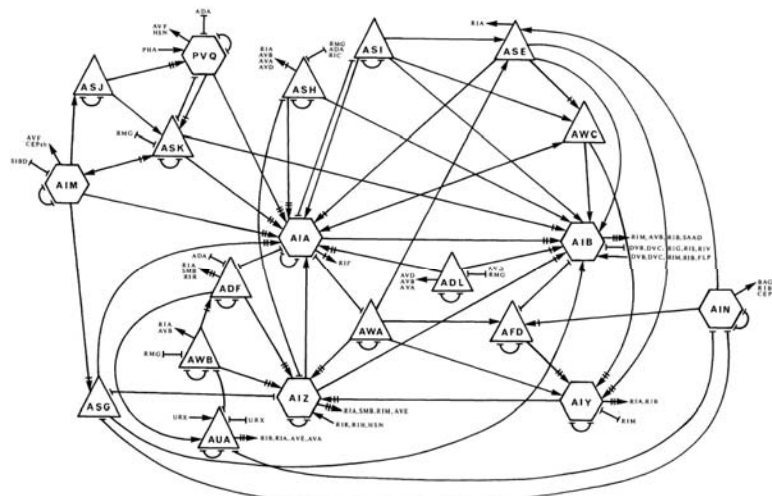
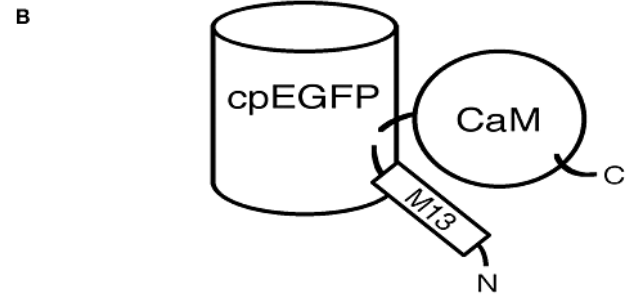
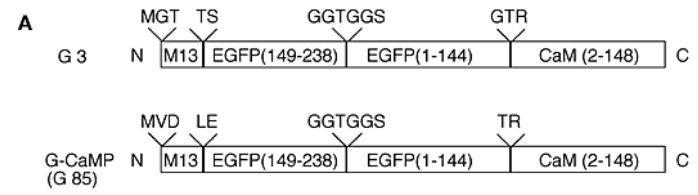
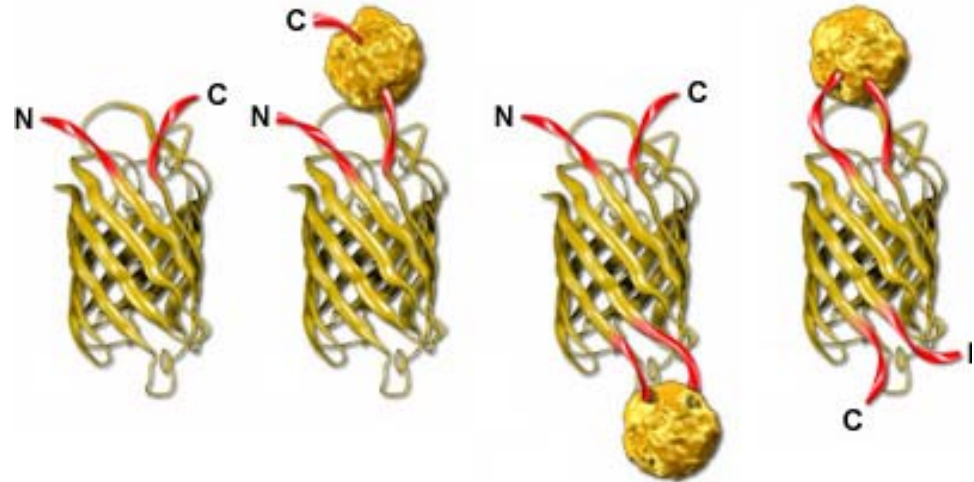
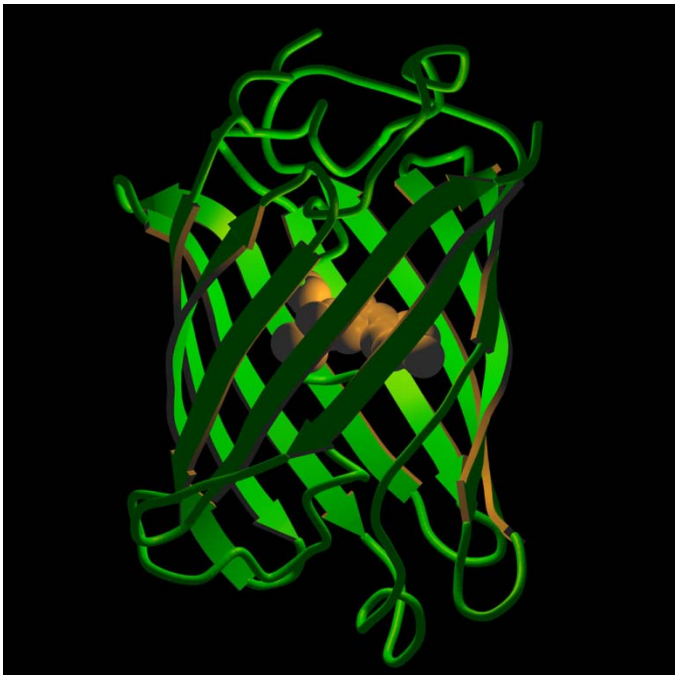
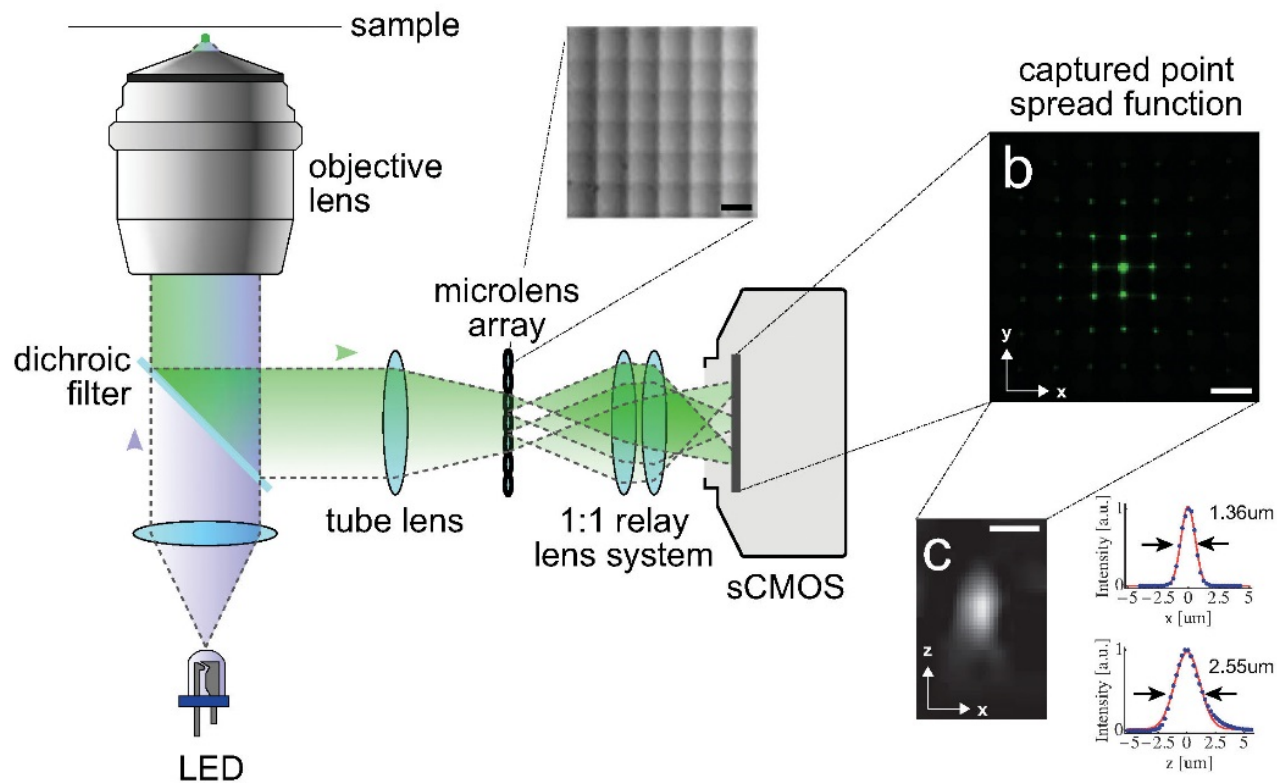


FIGURE 21. (a) Circuitry associated with amphids.





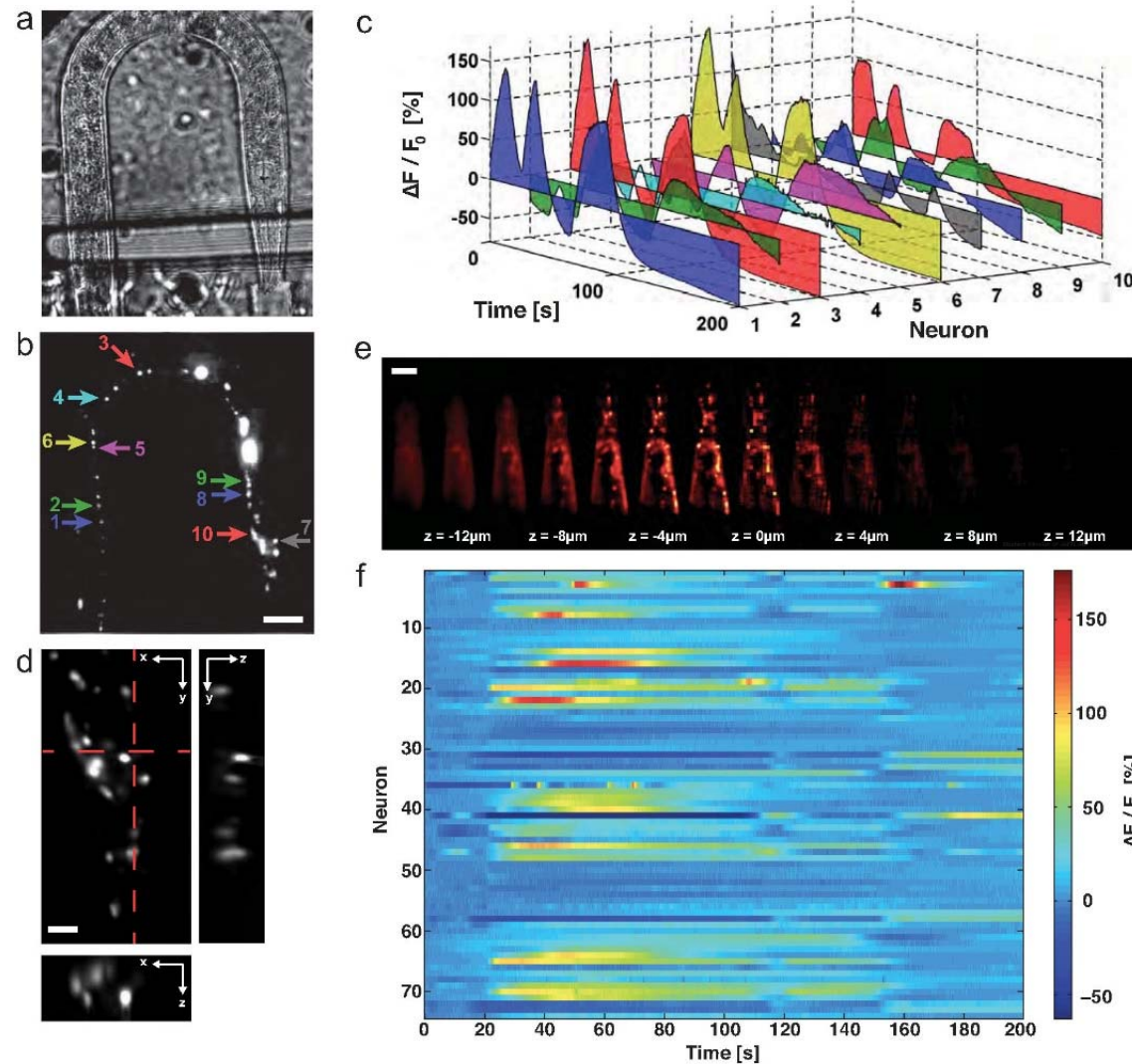
# Simultaneous, whole-animal, 3-D microscopy: light-field imaging



Prevedel\*, Yoon\*, et al. (2014) *Nature Methods*, advance online publication doi:10.1038/nmeth.2964.



# Simultaneous, whole-animal, 3-D imaging of neural activity (at 5-50 Hz)

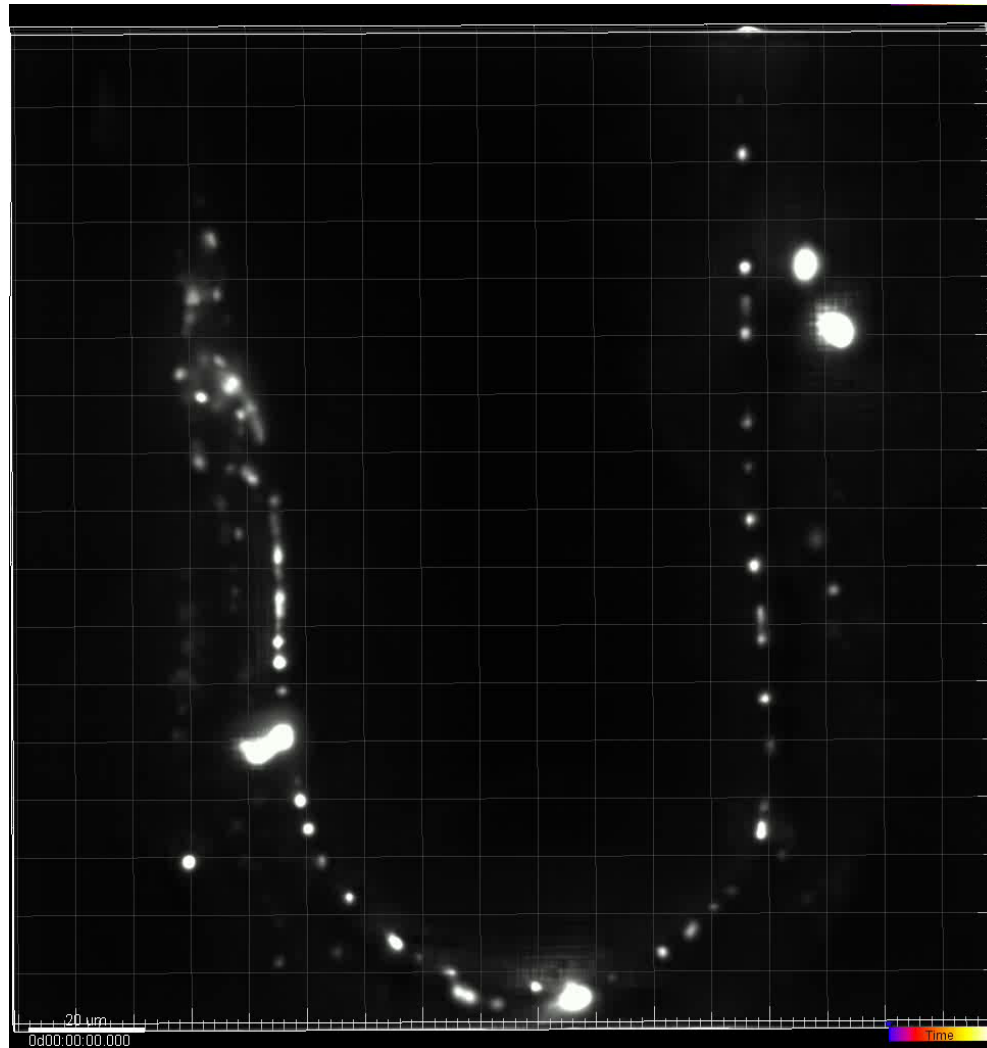


Prevedel\*, Yoon\*, et al. (2014) *Nature Methods*, advance online publication doi:10.1038/nmeth.2964.

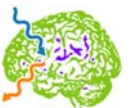




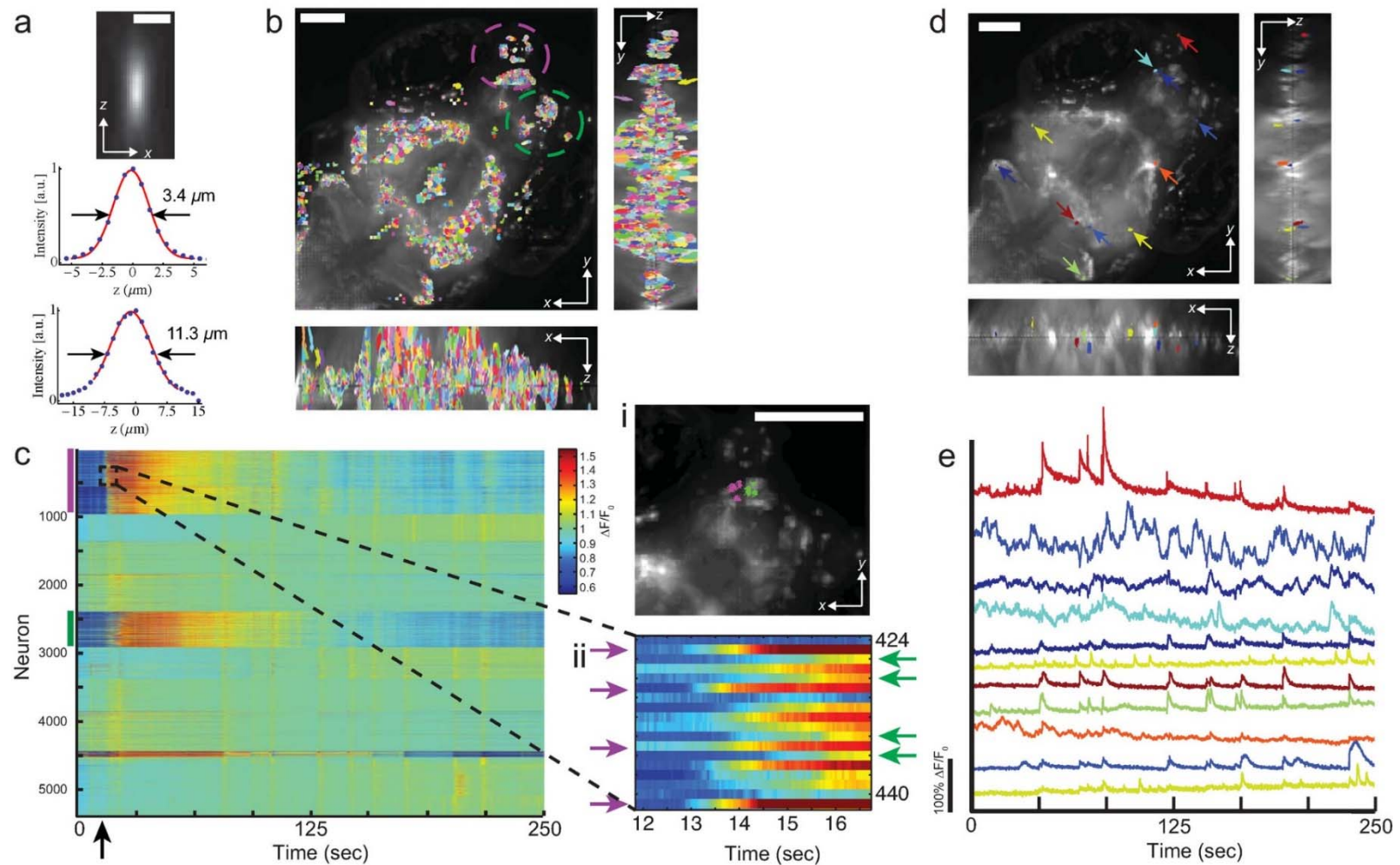
# Imaging neural activity throughout organism with known connectome



Prevedel\*, Yoon\*, et al. (2014) *Nature Methods*, advance online publication doi:10.1038/nmeth.2964.



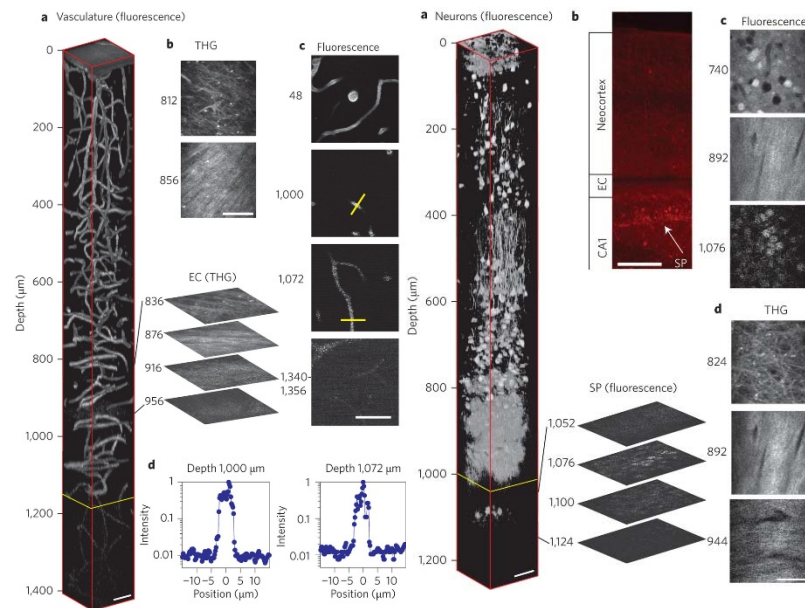
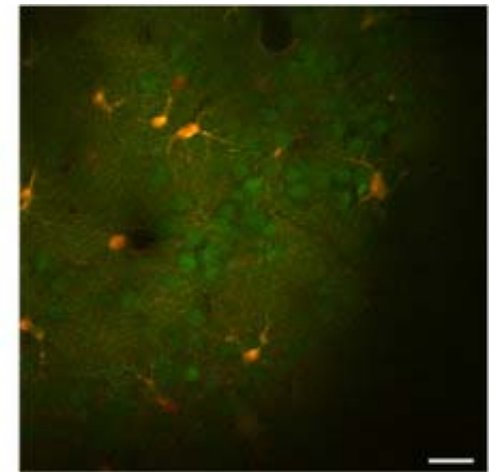
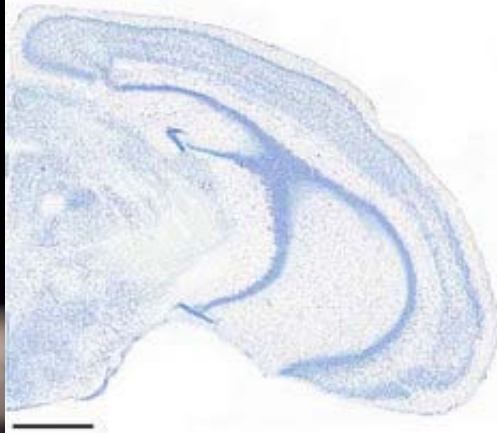
# Imaging zebrafish neural activity in 3-D (at 20 Hz, below)



Prevedel\*, Yoon\*, et al. (2014) *Nature Methods*, advance online publication doi:10.1038/nmeth.2964.



# The world's smallest mammal: towards whole-organism functional imaging



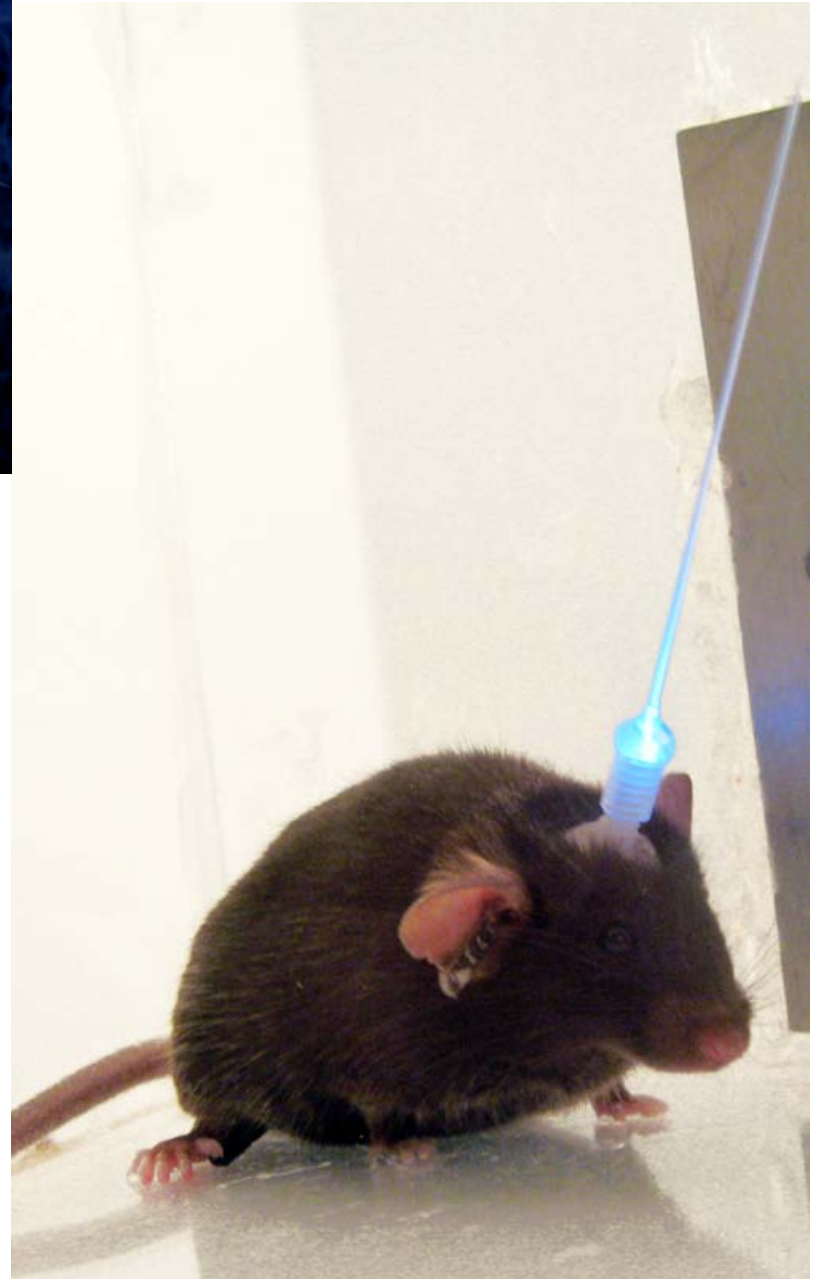
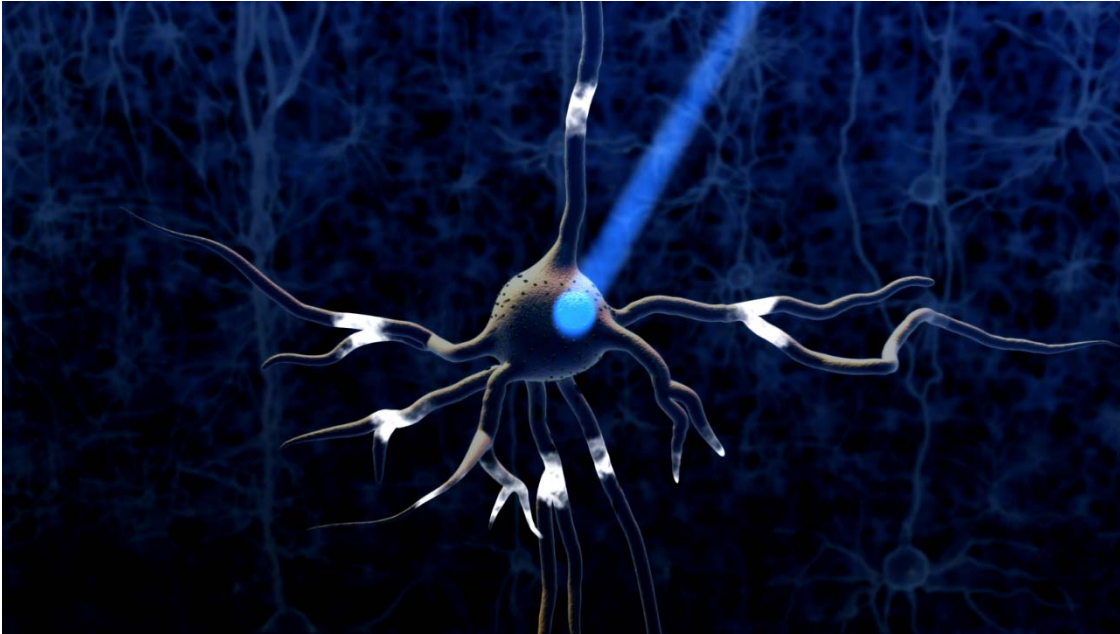
Michael Brecht, Ian Wickersham,  
Susan Erdman

<http://www.nature.com/nphoton/journal/v7/n3/abs/nphoton.2012.336.html>



# Can we understand causally how neurons function in circuits?



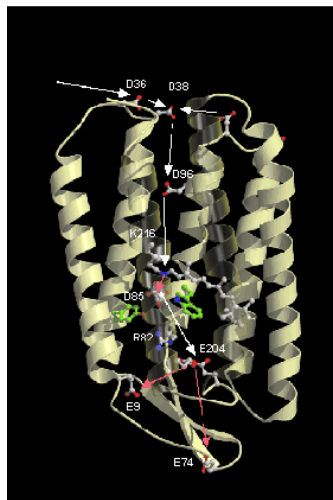




## Bacteriorhodopsins: Light-driven proton pumps



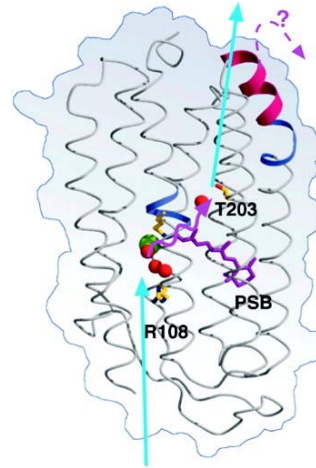
<http://www.genome.duke.edu/genomelife/2011/03/systems-under-stress/>



[http://www.biochem.mpg.de/523002/Protein\\_BR](http://www.biochem.mpg.de/523002/Protein_BR)

D. Oesterhelt and W. Stoeckenius (1971) Rhodopsin-like Protein from the Purple Membrane of Halobacterium halobium. Nature New Biology 233:149-152.

## Halorhodopsins: Light-driven chloride pumps



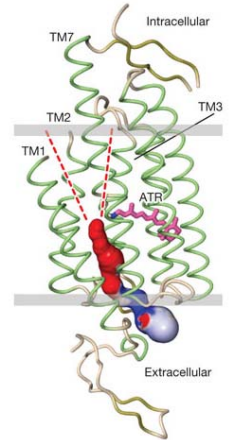
<http://www.sciencemag.org/content/288/5470/1390.full>

Matsuno-Yagi A, Mukohata Y (1977) Two possible roles of bacteriorhodopsin; a comparative study of strains of Halobacterium halobium differing in pigmentation. Biochem Biophys Res Commun 78:237-43.

Matsuno-Yagi A, Mukohata Y (1980) ATP synthesis linked to lightdependent proton uptake in a rad mutant strain of Halobacterium lacking bacteriorhodopsin. Arch Biochem Biophys, 199:297-303.

Schobert B, Lanyi JK (1982) Halorhodopsin is a light-driven chloride pump. J Biol Chem, 257:10306-13.

## Channelrhodopsins: Light-driven cation channels



<http://starcentral.mbl.edu/microscope/portal.php?pagetitle=assetfactsheet&imageid=3245>

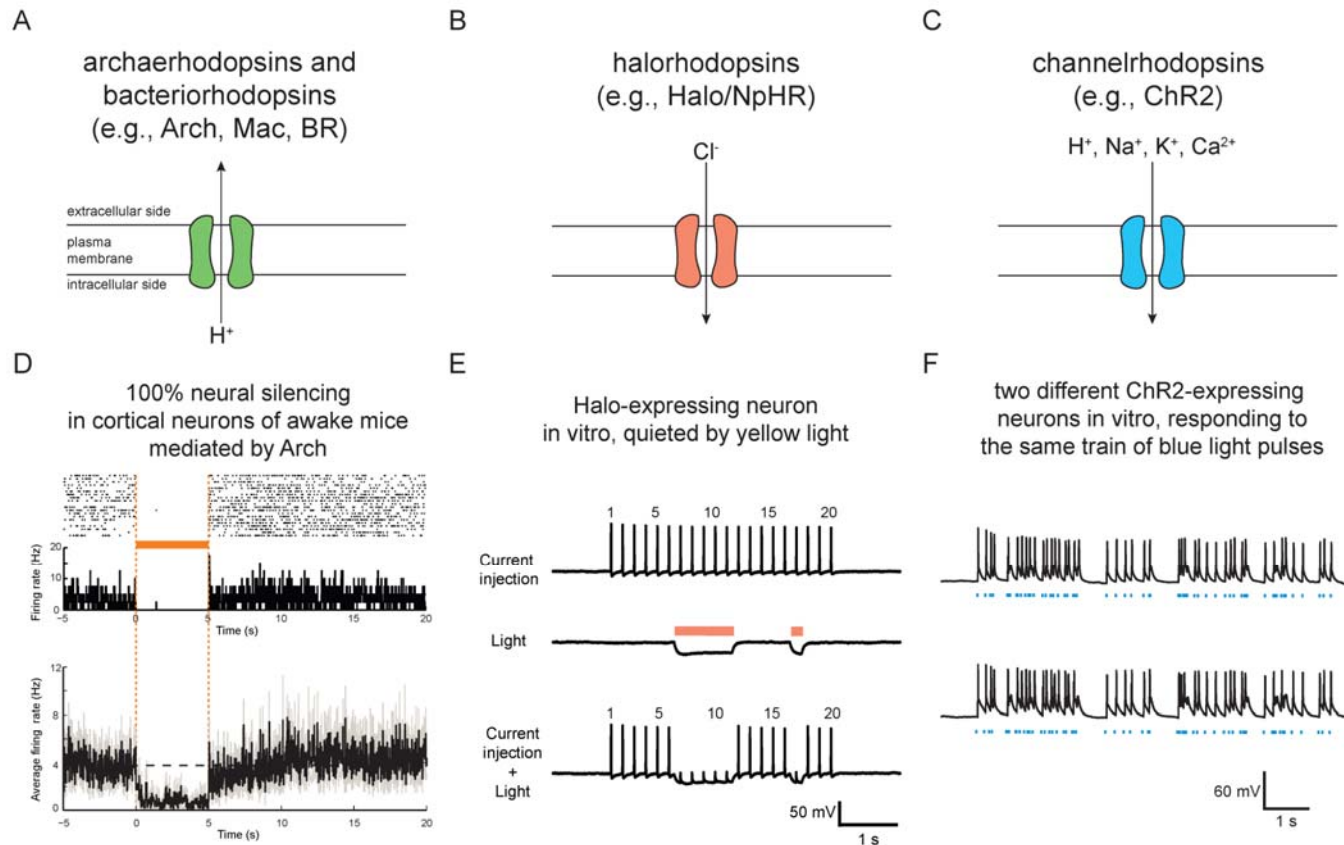
<http://www.nature.com/nature/journal/v482/n7385/full/nature10870.html>

Nagel G, Ollig D, Fuhrmann M, Kateriya S, Musti AM, Bamberg E, Hegemann P (2002) Channelrhodopsin-1: a light-gated proton channel in green algae. Science, 296:2395-8.

Nagel G, Szellas T, Huhn W, Kateriya S, Adeishvili N, Berthold P, Ollig D, Hegemann P, Bamberg E (2003) Channelrhodopsin-2, a directly light-gated cation-selective membrane channel. Proc Natl Acad Sci U S A, 100:13940-5.



# Three major optogenetic molecule classes: microbial opsins, seven-transmembrane proteins, binding endogenous all-trans-retinal





# Targeting different neurons of the brain in genetic model organisms, and beyond

## Lentiviruses and adeno-associated viruses

Have **intrinsic** tropism for certain cell types (e.g., lenti – excitatory neurons of the cortex)

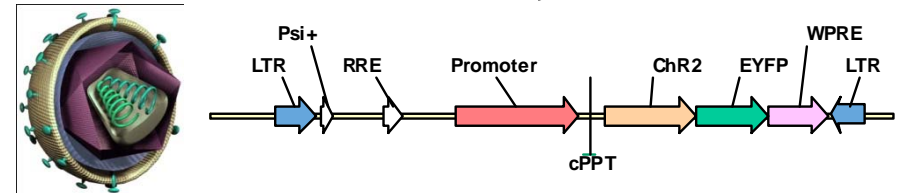
Can tune the **promoter**: synapsin pan neuronal, CaMKII excitatory, TH/dopamine, GAD, SOM, CCK, ...

AAV serotypes – AAV8, 5, 2, 9, ...

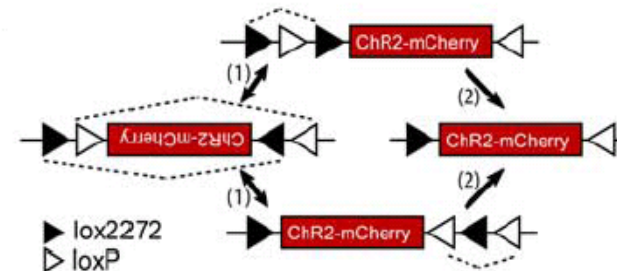
Lots of **Cre recombinase expressing mice** (e.g dopamine, serotonin, parvalbumin, etc.)

Administer a **floxed and reversed** opsin AAV into such a mouse, and the opsin will be flipped around into the correct direction

Takes **2-3 weeks** to express after injection; electroporation and other methods may be of use



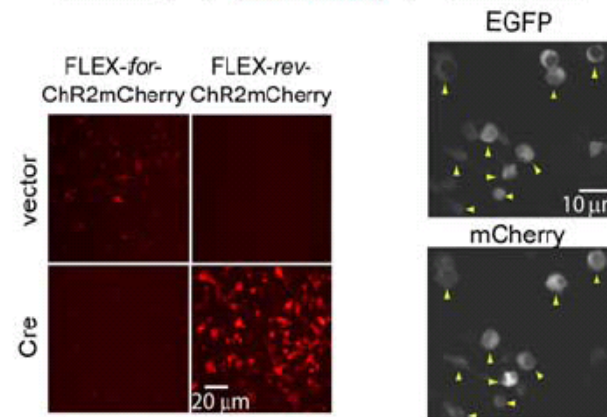
Atasoy et al., 2008



FLEX-for-ChR2mCherry: "on→off"



FLEX-rev-ChR2mCherry: "off→on"



# Rules of thumb for blue/green/yellow light

200 mW/mm<sup>2</sup> is good **irradiance** to shoot for (higher okay for brief neural activations; lower should be considered for long duration silencings; molecules are sensitive to 0.1-10 mW/mm<sup>2</sup>)

50 micron **fibers** = can easily go into tetrode drives

100-200 micron fibers = stiff enough to go into brain

400-800 micron fibers = for specialized uses

200 micron fiber, 200 mW/mm<sup>2</sup> affects **~1 mm<sup>3</sup>** of tissue

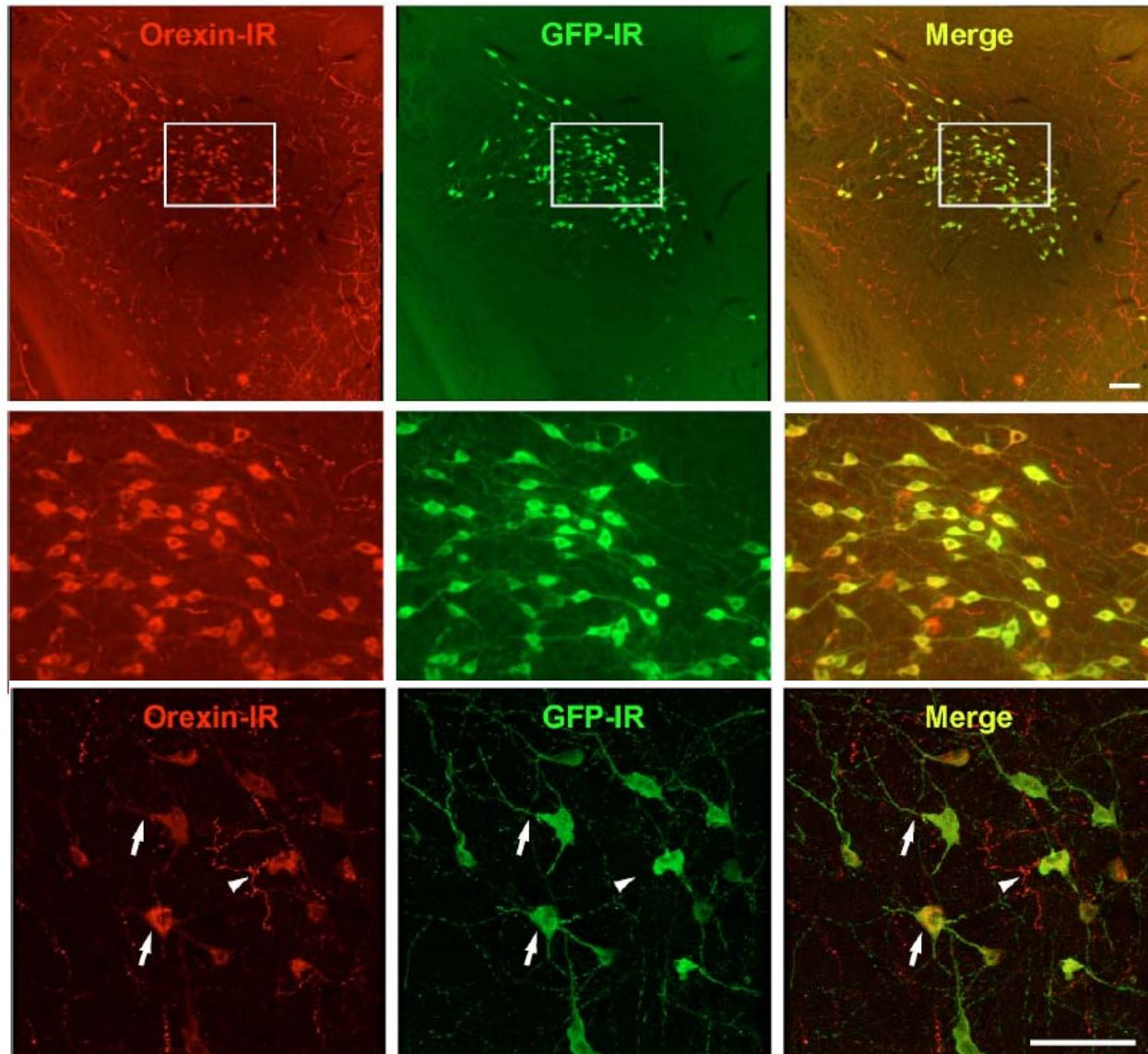
**Close** to fiber tip: light goes forward

Beyond a **scattering** length (~50-100 microns), starts to look spherical (power falls off as  $1/r^2$ )

Beyond the **absorbance** length (~500-1000 microns), falls off exponentially



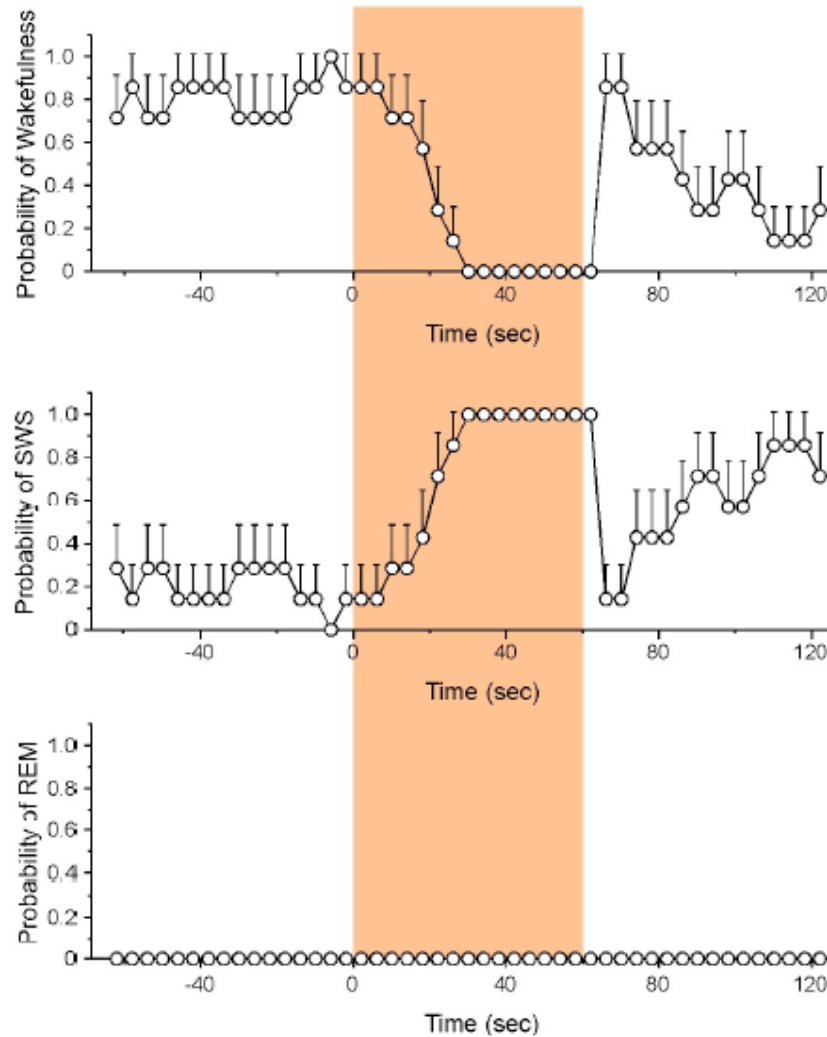
## Transgenic mice expressing original-N. pharaonis halorhodopsin, tagged with GFP, in hypocretin neurons



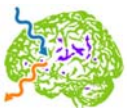
Tsunematsu et al. (2011) *Journal of Neuroscience* 31(29): 10529-10539.



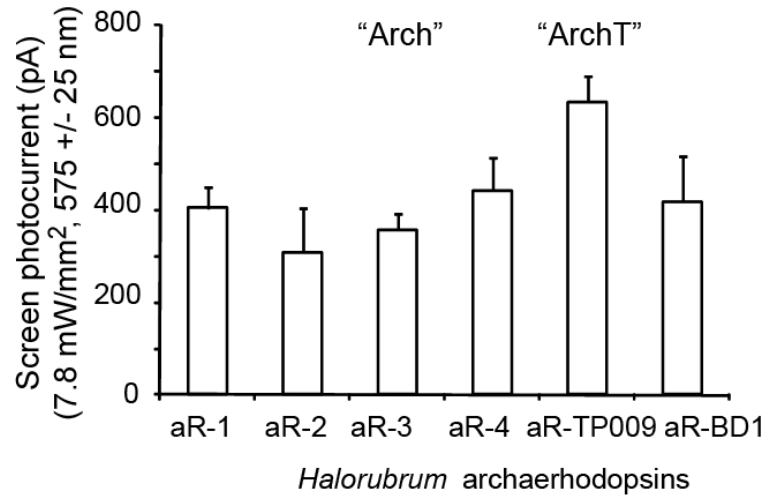
# Light silences the neurons, resulting in slow-wave sleep



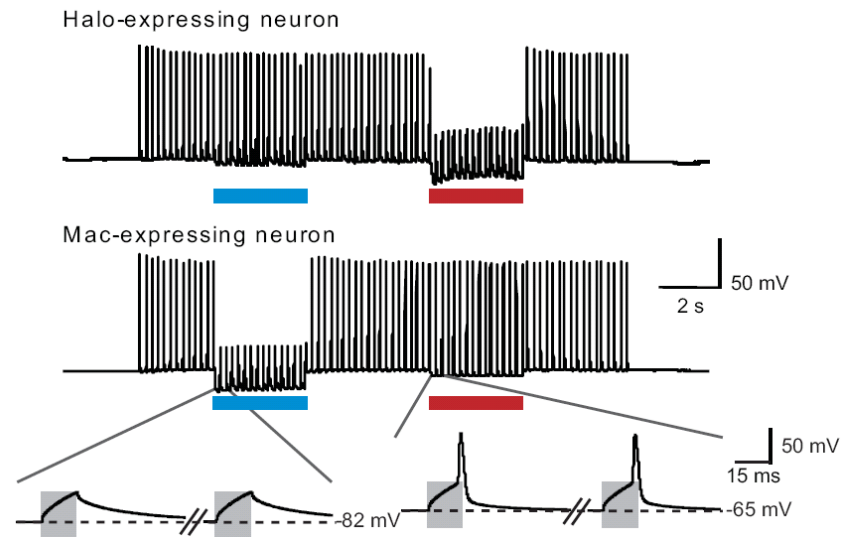
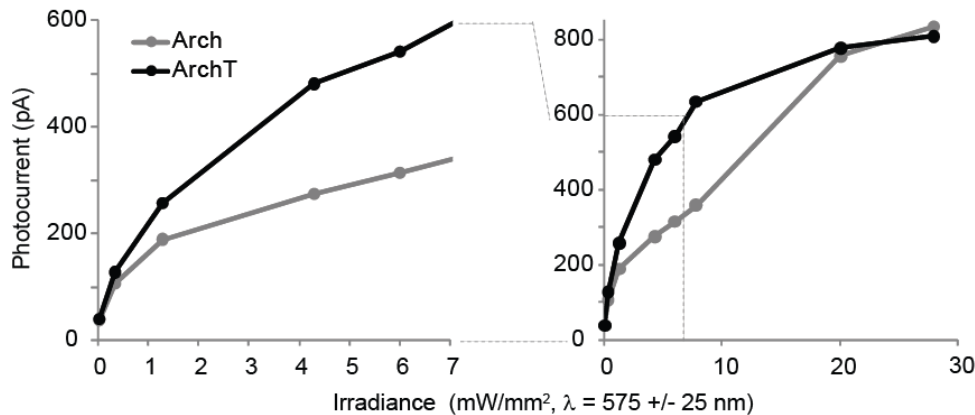
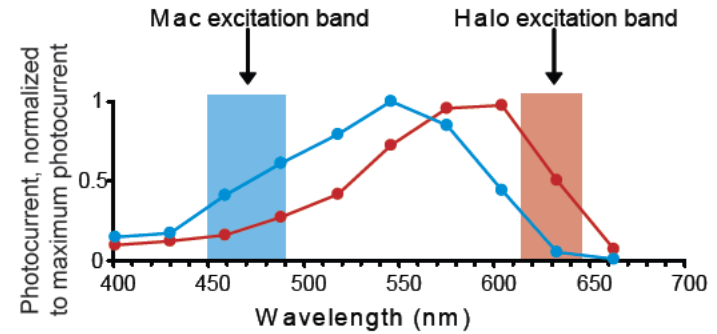
Tsunematsu et al. (2011) *Journal of Neuroscience* 31(29): 10529-10539.



**Search locally in genomic space:  
ArchT, higher light sensitivity relative  
of Arch**



**Search broadly in genomic space:  
Mac, blueshifted relative to all other  
silencers**

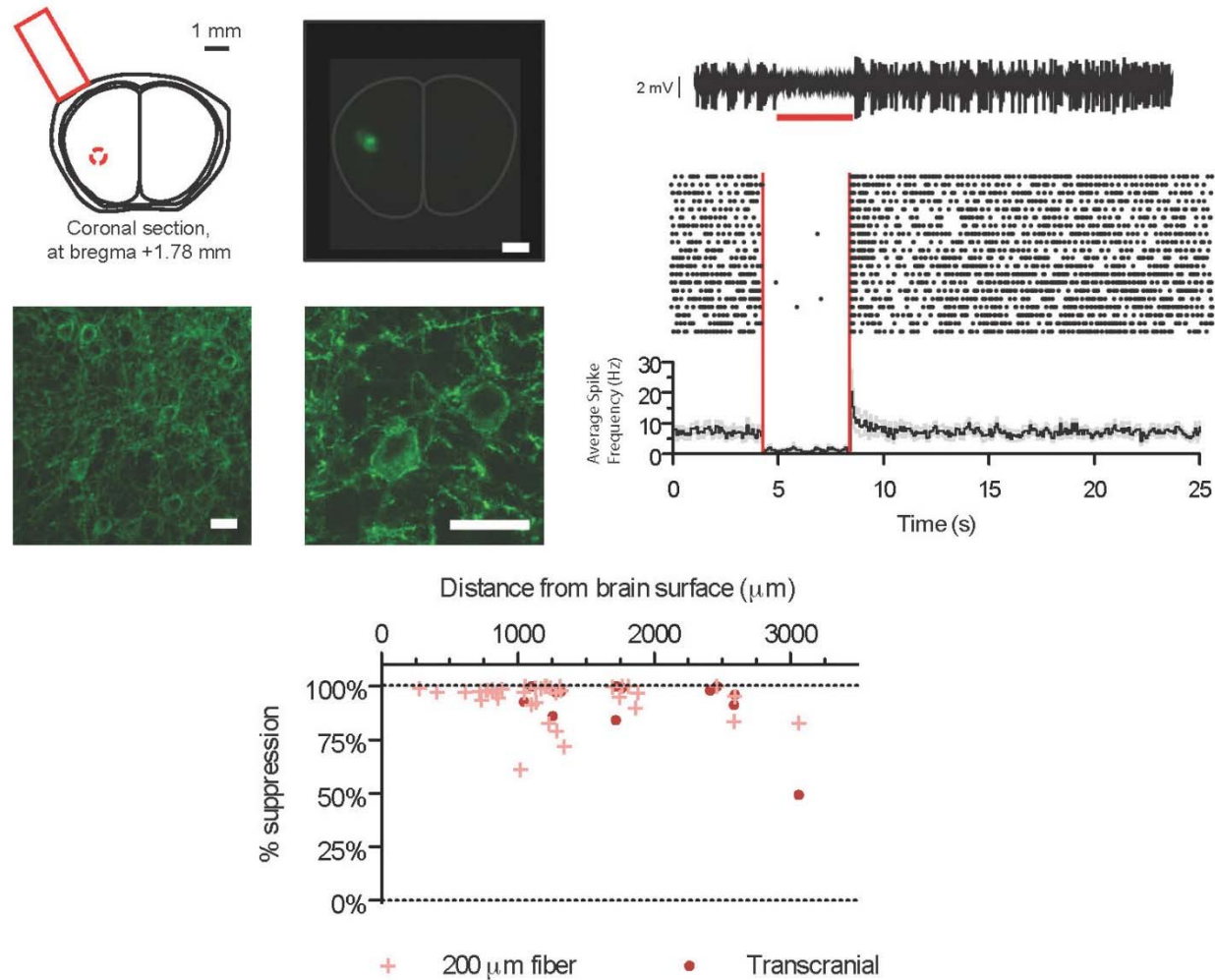


**Han\*, Chow\*, et al. (2011) *Frontiers in Systems Neuroscience* 5:18.**

**Chow\*, Han\*, et al. (2010) *Nature* 463:98-102.**



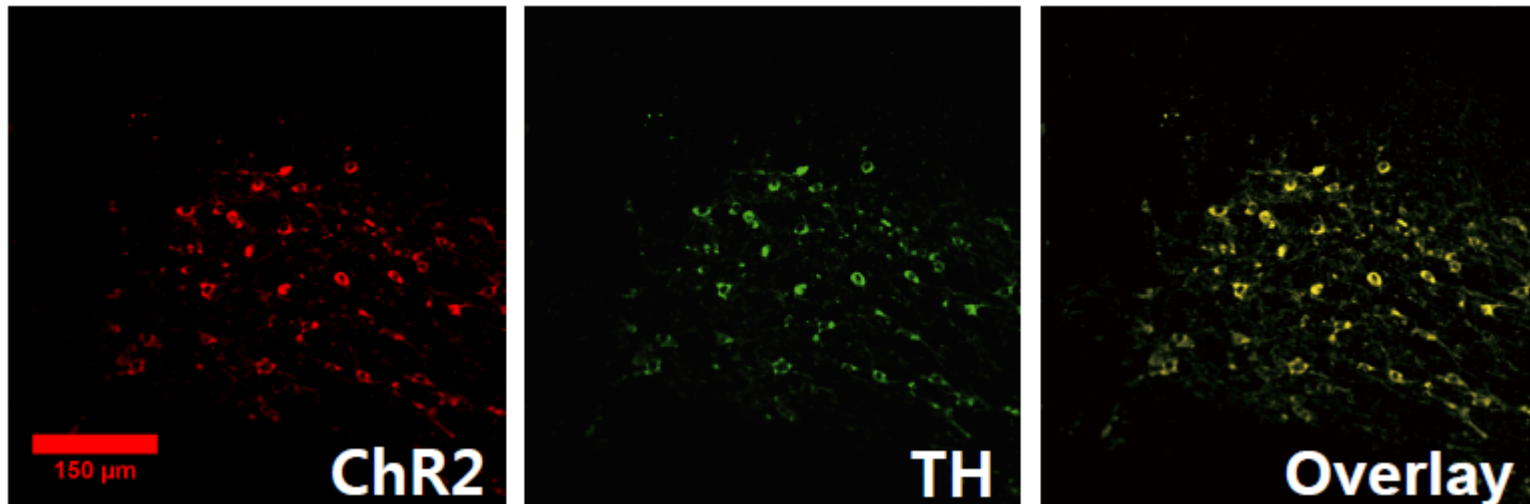
# Noninvasive optogenetic neural silencing: Jaws



Chuong et al. (2014) *Nature Neuroscience*, accepted.



# DAT-Cre + AAV-FLEX-ChR2-tdTomato



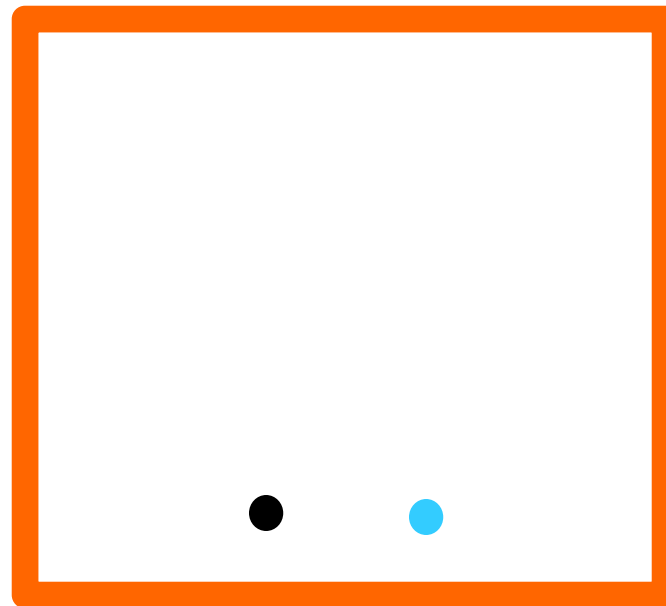
# Finding circuits in the brain that can mediate reward

## Dopamine

### neurons:

implicated in reward and addiction, but largely through pharmacological and electrical means

Is a **brief activation** of them sufficient to drive reward?



**no light  
stimulation**

**light  
stimulation**



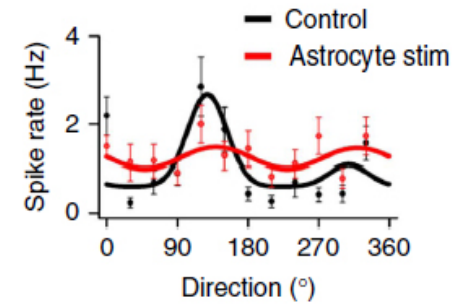
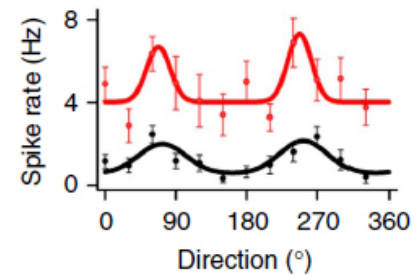
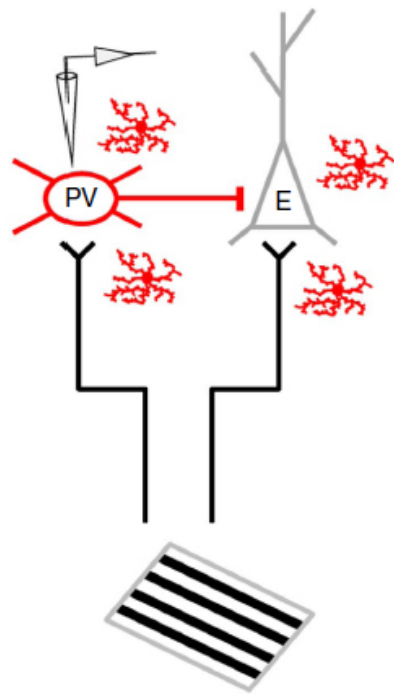




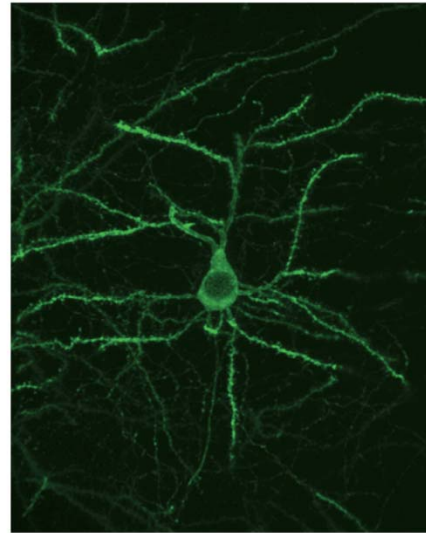
Kim et al. (2012) *PLoS One* 7(4):e33612



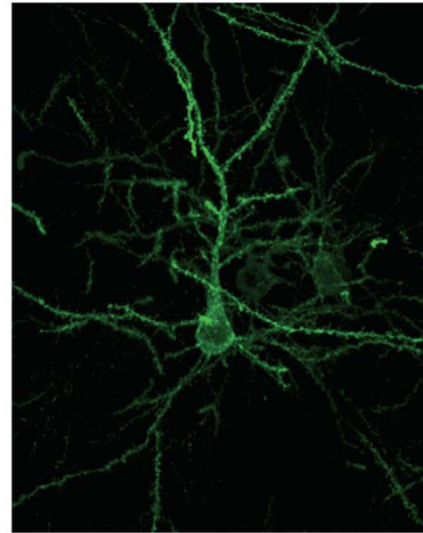
# Optogenetic activation of glia can change neural codes



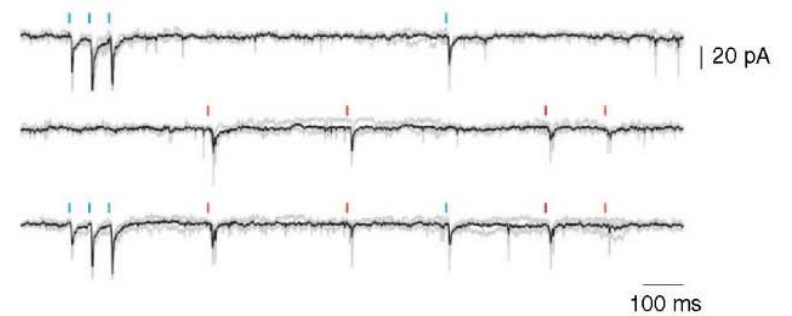
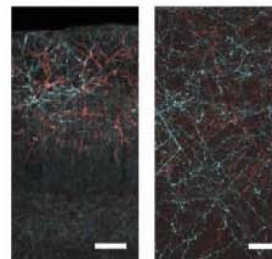
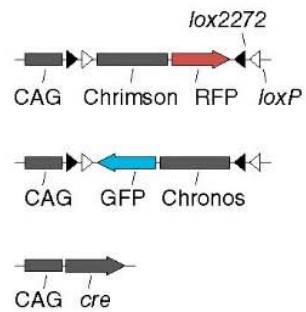
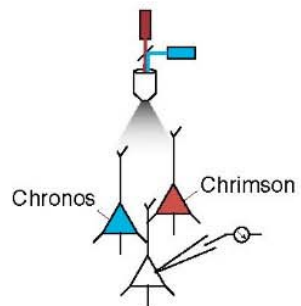
# Chronos and Chrimson together



Chronos



Chrimson



Klapoetke et al. (2014) *Nature Methods* 11:338–346.



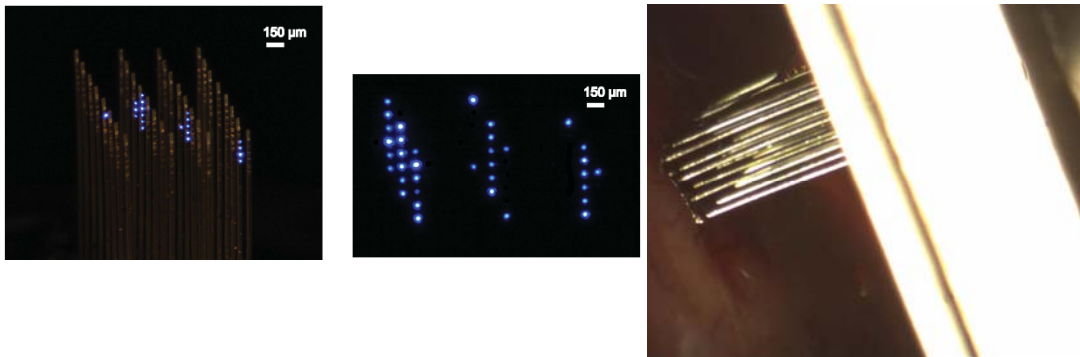
# Accessory strategies for a diversity of systems

## Wireless, multisite optogenetics



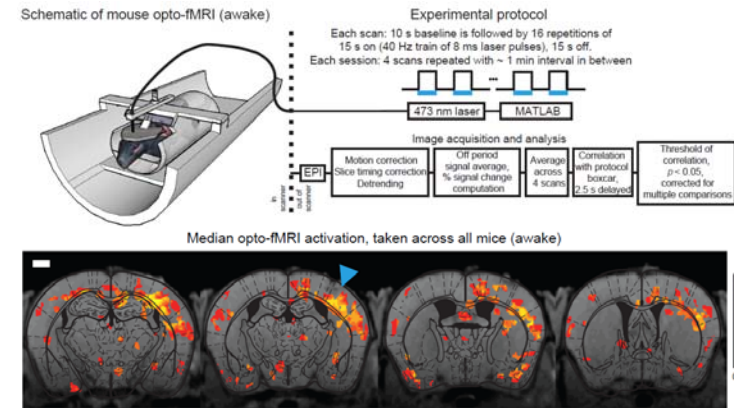
Wentz et al. (2011) *Journal of Neural Engineering* 8(4): 046021, commercialized by Kendall Research Systems, Inc. (ESB has no financial affiliation); Bernstein and Boyden (2011) *Trends in Cognitive Sciences* 15(12):592-600; Bernstein et al. (2011) *Current Opinion in Neurobiology* 22(1):61-71.

## 3-D optogenetic control



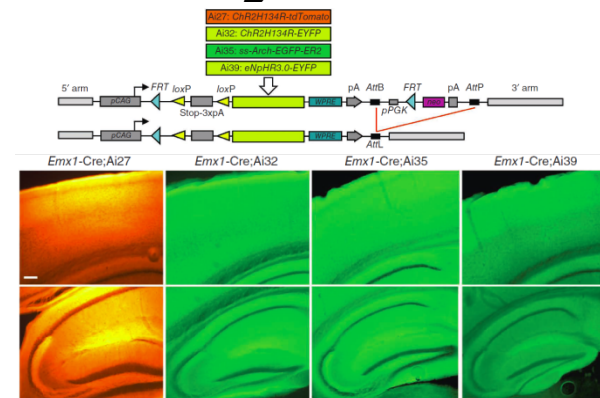
Zorzos et al. (2010) *Optics Letters* 35(24):4133-5; Zorzos et al. (2012) *Optics Letters* 37(23):4841-4843

## Opto-fMRI



Desai et al. (2011) *Journal of Neurophysiology* 105(3):1393-405; Kahn et al. (2011) *Journal of Neuroscience* 31(42):15086-15091.

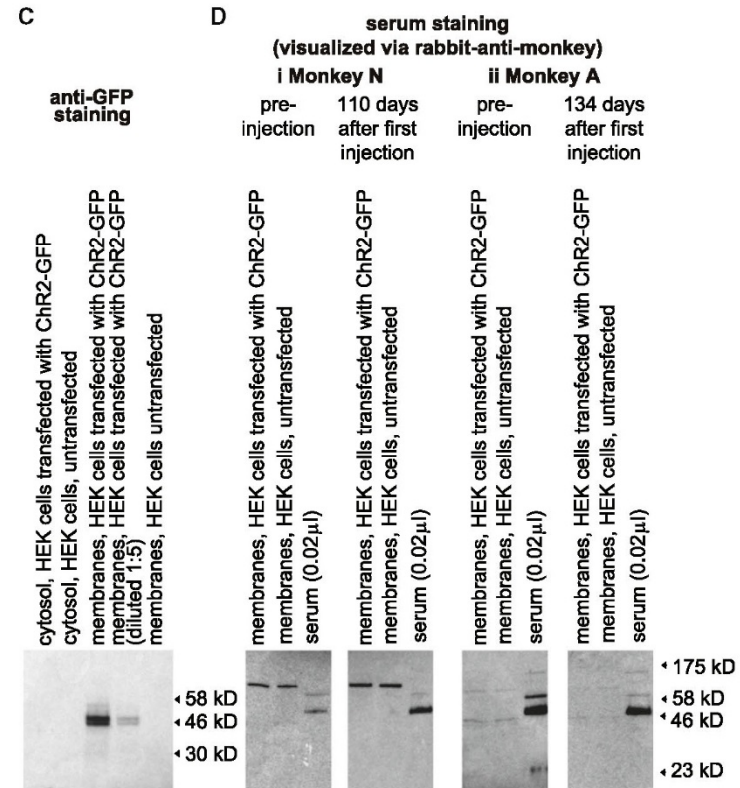
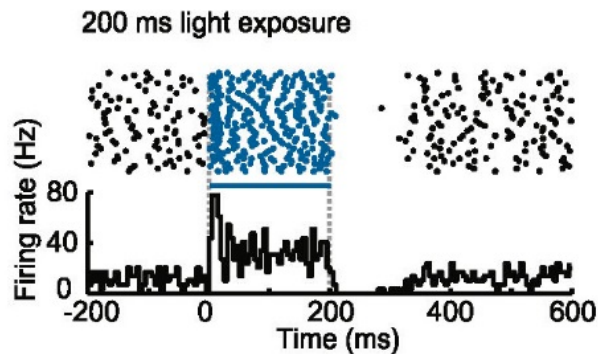
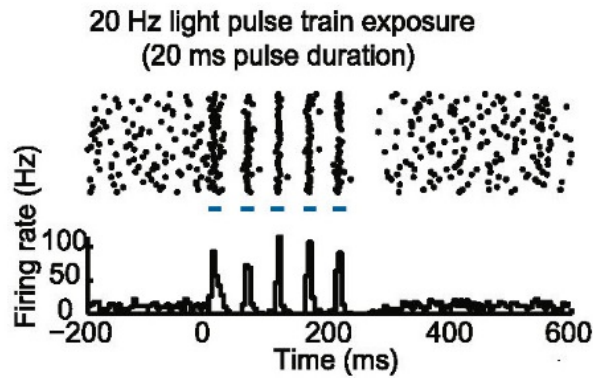
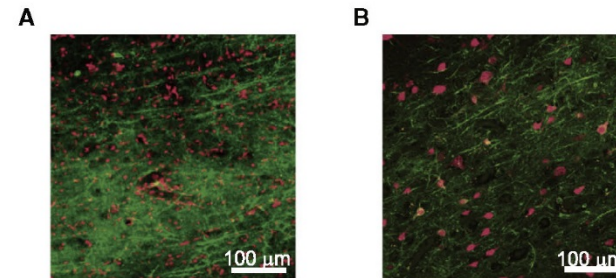
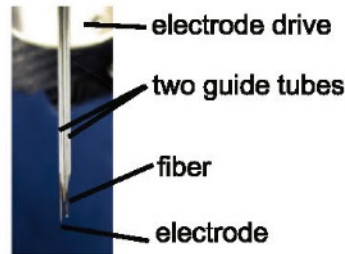
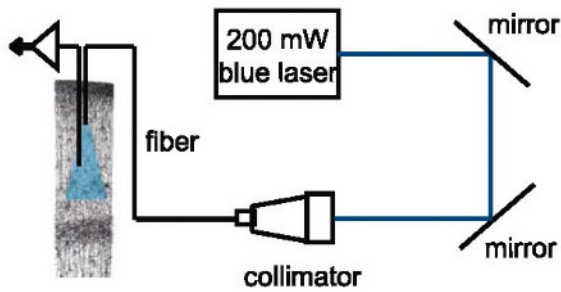
## Transgenic mice



Madisen et al. (2012) *Nature Neuroscience* 15(5):793-802.



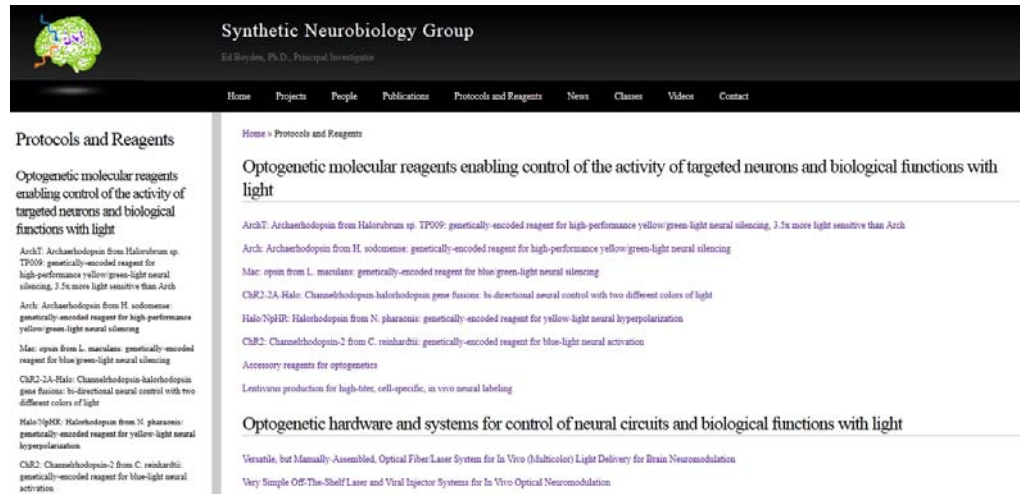
# Primate optogenetics



Han et al. (2009) *Neuron* 62(2):191-198.



# Syntheticneurobiology.org



**Synthetic Neurobiology Group**  
Ed Ruthven, Ph.D., Principal Investigator

Home Projects People Publications Protocols and Reagents News Classes Videos Contact

## Protocols and Reagents

Home » Protocols and Reagents

### Optogenetic molecular reagents enabling control of the activity of targeted neurons and biological functions with light

ArchT: Archaelhodopsin from Halorubrum sp. TP009: genetically-encoded reagent for high-performance yellow/green-light neural silencing, 2.5x more light sensitive than Arch

Arch: Archaelhodopsin from H. salomonense: genetically-encoded reagent for high-performance yellow/green-light neural silencing

Mac: opsin from L. maculans: genetically-encoded reagent for blue/green-light neural silencing

ChR2-2A-Halo: Channelhodopsin-halorhodopsin gene fusions: bi-directional neural control with two different colors of light

Halo2yHR: Halorhodopsin from N. pharaonis: genetically-encoded reagent for yellow light neural hyperpolarization

ChR2: Channelhodopsin-2 from C. reinhardtii: genetically-encoded reagent for blue-light neural activation

Accessory reagents for optogenetics

Lentiviral production for high-titer, cell-specific, in vivo neural labeling

### Optogenetic hardware and systems for control of neural circuits and biological functions with light

Versatile, but Manually-Assembled, Optical Fiber/Laser System for In Vivo (Multicolor) Light Delivery for Brain Neurostimulation

Very Simple Off-The-Shelf Laser and Viral Injector Systems for In Vivo Optical Neurostimulation

Tools distributed to >1000 labs worldwide

- Addgene, DNA
- UNC (Lori Nisi), viruses
- Allen Institute (Hongkui Zeng), floxed-stop transgenics
- Host visitors (1-2x/week) to teach procedures



### Graduate Students, Postdocs, Staff

Adam Marblestone  
Aimei Yang  
Amy Chuong  
Annabelle Singer  
Anthony Zorzos  
Asmamaw Wassie  
Brian Allen  
Caroline Moore-Kochlacs  
Christian Wentz  
Changyang Linghu  
Daniel Martin-Alarcon  
Daniel Schmidt  
Desiree Dudley  
Erica Jung  
Fei Chen  
Fumi Yoshida  
Giovanni Talei Franzesi  
Guangyu Xu  
Harbaljit Sohal  
Ho-Jun Suk  
Ishan Gupta  
Jae-Byum Chang  
Jake Bernstein  
Jorg Scholvin  
Jun Deguchi  
Justin Kinney  
Kate Adamala  
Kiryl Piatkevich  
Kris Payer  
Leah Acker  
Lisa Lieberson  
Manos Karagiannis  
Mike Henninger  
Moshe Ben-Ezra  
Nathan Klapoetke  
Nikita Pak  
Nir Grossman  
Or Shemesh  
Paul Tillberg  
Ru Wang  
Suhasa Kodandaramaiah  
Yongxin Zhao  
Young Gyu Yoon

### Alumni

Alexander Guerra, Alex Rodriguez, Allison Dobry, Ash Turza, August Dietrich, Barbara Barry, Brian Chow (U Penn), Claire Ahn, Nate Greenslit (Harvard), Ian Wickersham (MIT), Ilya Kolb, Jenna Sternberg, Kyungman Kim, Masaaki Ogawa (NIPS), Masahiro Yamaguchi, Mike Baratta, Mingjie Li, Rachel Bandler, Scott Arfin, Stephanie Chan, Tania Morimoto, Tim Buschman (Princeton), Victoria Wang, Xiaofeng Qian, Xue Han (BU), Yongku Cho (U. Conn.)

### Undergraduate Students

Alexander Clifton, Bara Badwan, Denis Bozic, Deniz Aksel, Eunice Wu, Justine Cheng, Melina Tsitsiklis, Rebecca Luoh, Rita Ainane, Sean Batir, Sunanda Sharma

# Synthetic Neurobiology Group

<http://syntheticneurobiology.org/>

### Funding

Allen Institute for Brain Science; AT&T; Bahaa Hariri; Benesse Corporation; Jerry and Marge Burnett; DARPA Living Foundries Program HR0011-12-C-0068; DARPA HR0011-11-14-0004; Department of Defense CDMRP PTSD Program; Google; Harvard/MIT Joint Grants Program in Basic Neuroscience; Human Frontiers Science Program; IET A. F. Harvey Prize; Joyce and Jeremy Wertheimer; Lincoln Labs Campus Collaboration Award; MIT Alumni Class Funds; MIT Intelligence Initiative; MIT McGovern Institute and McGovern Institute Neurotechnology (MINT) Program; MIT Media Lab and Media Lab Consortia; MIT Mind-Machine Project; MIT Neurotechnology Fund (& its generous donors); NARSAD; New York Stem Cell Foundation-Robertson Investigator Award; NIH Director's Pioneer Award 1DP1NS087724 and New Innovator Award 1DP2OD002002, NIH EUREKA Awards 1R01NS087950 and 1R01NS075421, NIH Transformative Awards 1R01MH103910 and 1R01GM104948, NIH Single Cell Grant 1R01EY023173, and NIH Grants 1R01DA029639, 1R43NS070453, 1RC2DE020919, 1RC1MH088182, 2R44NS070453, and 1R01NS067199; NSF INSPIRE Award CBET 1344219, NSF CAREER Award CBET 1053233 and NSF Grants, EFR10835878, DMS0848804, and DMS1042134 (the Cognitive Rhythms Collaborative); Office of the Assistant Secretary of Defense for Research and Engineering; Paul Allen Distinguished Investigator in Neuroscience Award; Skolkovo Institute of Science and Technology; Alfred P. Sloan Foundation; Society for Neuroscience Research Award for Innovation in Neuroscience (RAIN); Stacy and Joel Hock; Synthetic Intelligence Project (& its generous donors); Wallace H. Coulter Foundation. Pre-MIT: Hertz Foundation, Helen Hay Whitney Foundation.

### Collaborators

**3-D Brain-building:** Utkan Demirci  
**Blindness:** Alan Horsager, Ben Matteo, Bill Hauswirth, Alapakkam Sampath

**C. elegans:** Ramesh Raskar, Alipasha Vaziri, Manuel Zimmer

**Microscopy:** Ramesh Raskar, Peter So, Alipasha Vaziri

**Neural modeling:** Nancy Kopell, Christoph Borger, Miles Whittington, Fiona LeBeau

**Neural recording:** Leaf labs, Konrad Kording, George Church

**Opsin engineering:** Gane Wong, Beijing Genomics Institute, Patrick Stern, Yingxi Lin, Ernst Bamberg, Michael Melkonian, Adam Cohen, Vivek Jayaraman, Martha Constantine-Paton

**Opto-fMRI:** Chris Moore, Randy Buckner, Itamar Kahn, Ann Graybiel, Nancy Kopell

**Optogenetic hardware:** Clif Fonstad, Rahul Sarpeshkar, Steve Wasserman, Ramesh Raskar, Joseph Jacobson, Ferro Solutions Inc., Kendall Research Systems

**Polymerase engineering:** George Church, Konrad Koering, Keith Tyo

**Primate work:** Bob Desimone, Ann Graybiel, Roderick Bronson, Wim Vanduffel, Bob Wurtz

**In vivo Robotics:** Craig Forest

**Transgenics:** Hongkui Zeng

